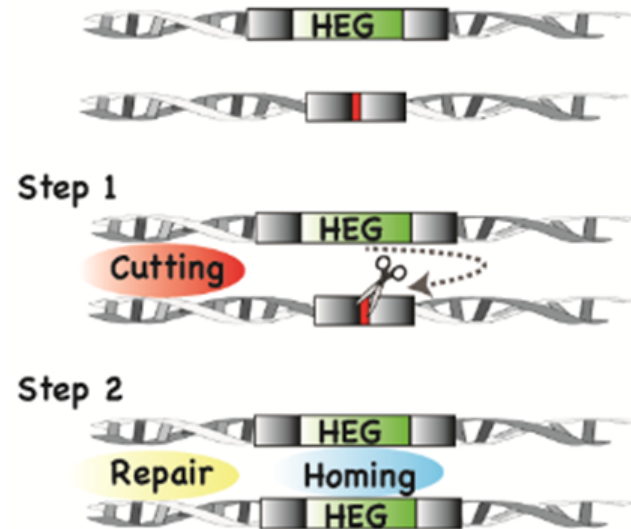
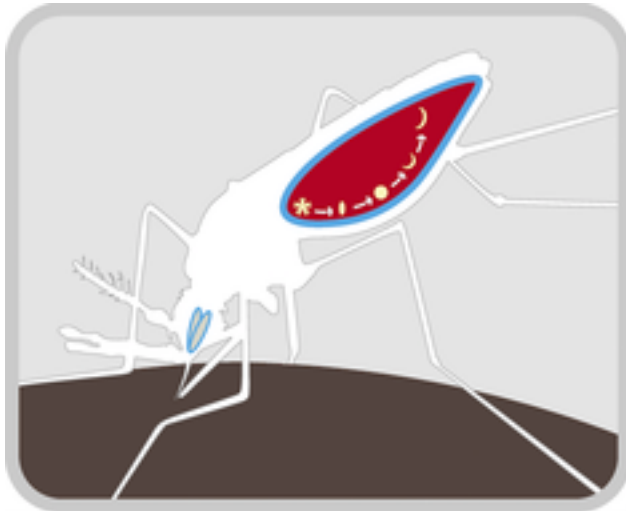


Harvard Medical School, 28<sup>th</sup> March 2018

# Can malaria, dengue and Zika be controlled by CRISPR-based gene drive and other novel tools?

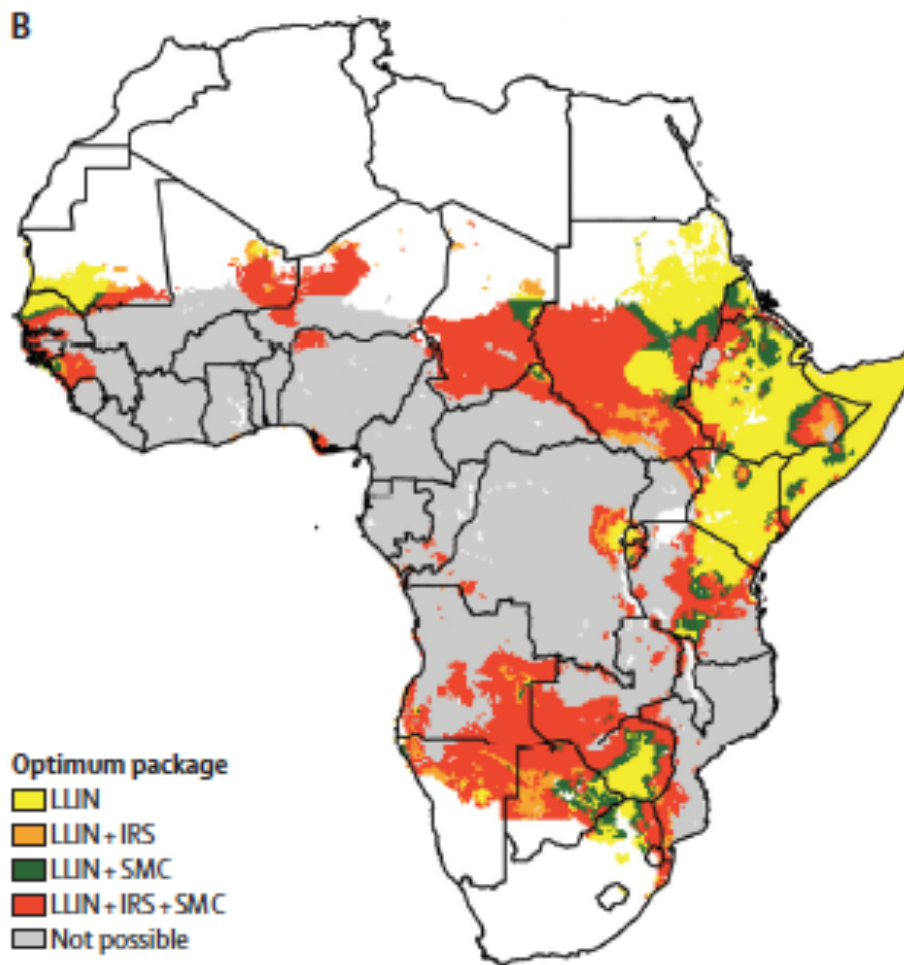
John M. Marshall

Divisions of Biostatistics & Epidemiology  
School of Public Health, University of California, Berkeley



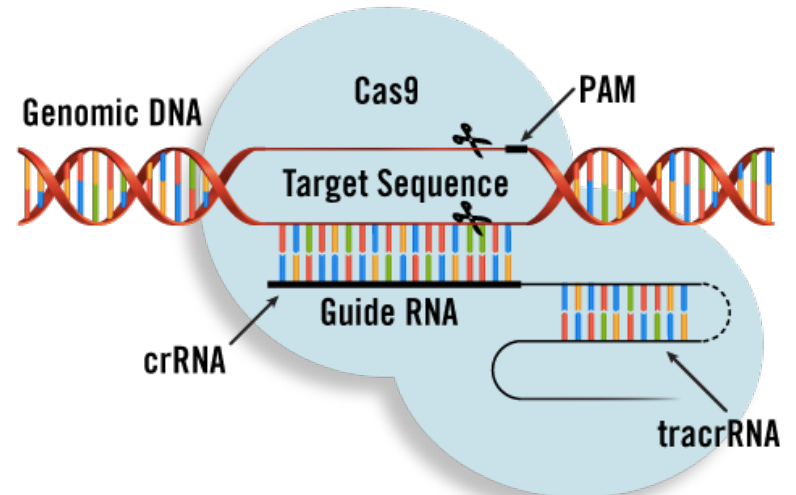
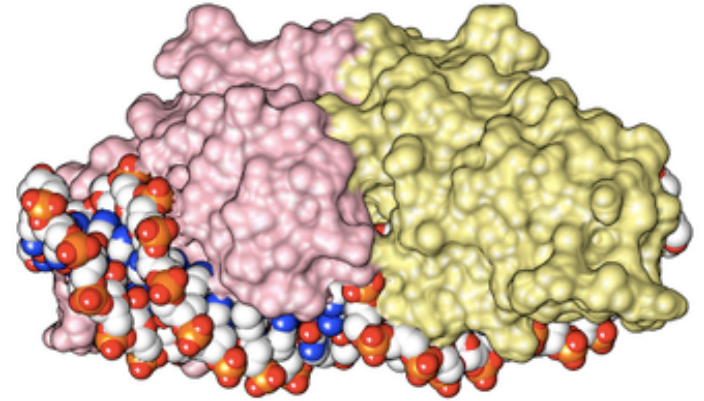
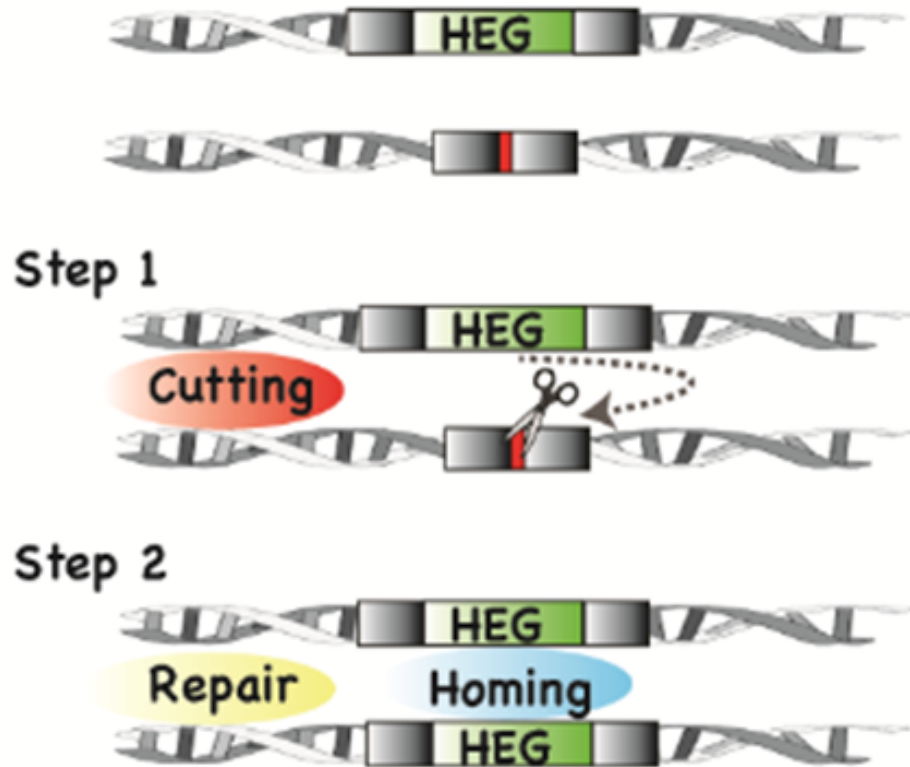
# Estimating the most efficient allocation of interventions to achieve reductions in *Plasmodium falciparum* malaria burden and transmission in Africa: a modelling study

Patrick GT Walker, Jamie T Griffin, Neil M Ferguson, Azra C Ghani



- Walker PGT, Griffin JT, Ghani AC (2016) Lancet Global Health 4: e474-e484.

# Homing-based gene drive systems



# Recent engineering successes

## The mutagenic chain reaction: A method for converting heterozygous to homozygous mutations

Valentino M. Gantz\* and Ethan Bier\*

The logo for ScienceExpress, featuring the word "Science" in a bold, sans-serif font and "Express" in a white, italicized, sans-serif font, both set against a black rectangular background.The logo for the Proceedings of the National Academy of Sciences (PNAS), consisting of the letters "PNAS" in a white, sans-serif font, oriented vertically on a blue rectangular background.

## Highly efficient Cas9-mediated gene drive for population modification of the malaria vector mosquito *Anopheles stephensi*

Valentino M. Gantz<sup>a,1</sup>, Nijole Jasinskiene<sup>b,1</sup>, Olga Tatarenkova<sup>b</sup>, Aniko Fazekas<sup>b</sup>, Vanessa M. Macias<sup>b</sup>, Ethan Bier<sup>a,2</sup>, and Anthony A. James<sup>b,c,2</sup>

## LETTERS

nature  
biotechnology

A CRISPR-Cas9 gene drive system targeting female reproduction in the malaria mosquito vector *Anopheles gambiae*

# Q1. Is it possible to perform a confined trial of a gene drive system?

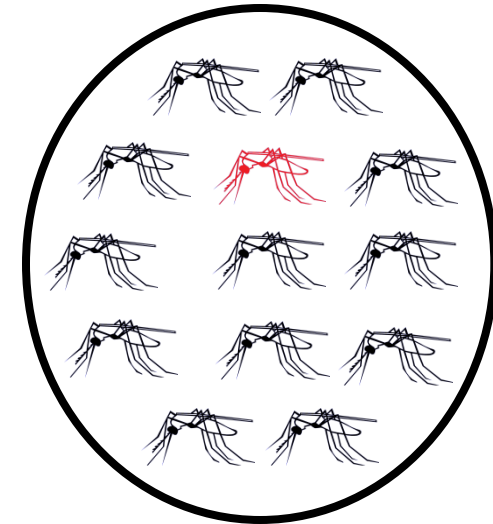
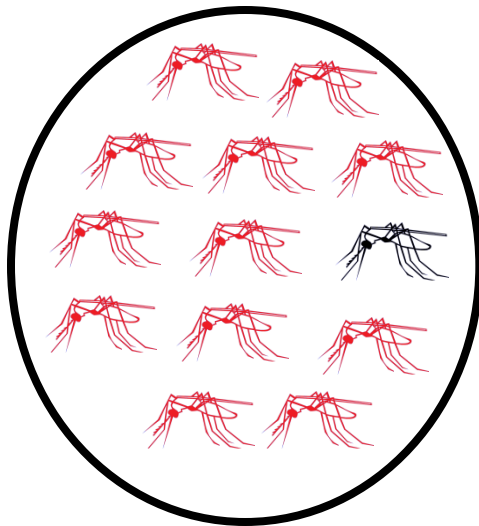


**Banambani, Mali**

7 km  
~1% / gen



**Doneguebougou, Mali**



# Perspectives of people in Mali toward genetically-modified mosquitoes for malaria control

John M Marshall\*<sup>1</sup>, Mahamoudou B Touré<sup>2</sup>, Mohamed M Traore<sup>2</sup>, Shannon Famenini<sup>3,4</sup> and Charles E Taylor<sup>3,4</sup>

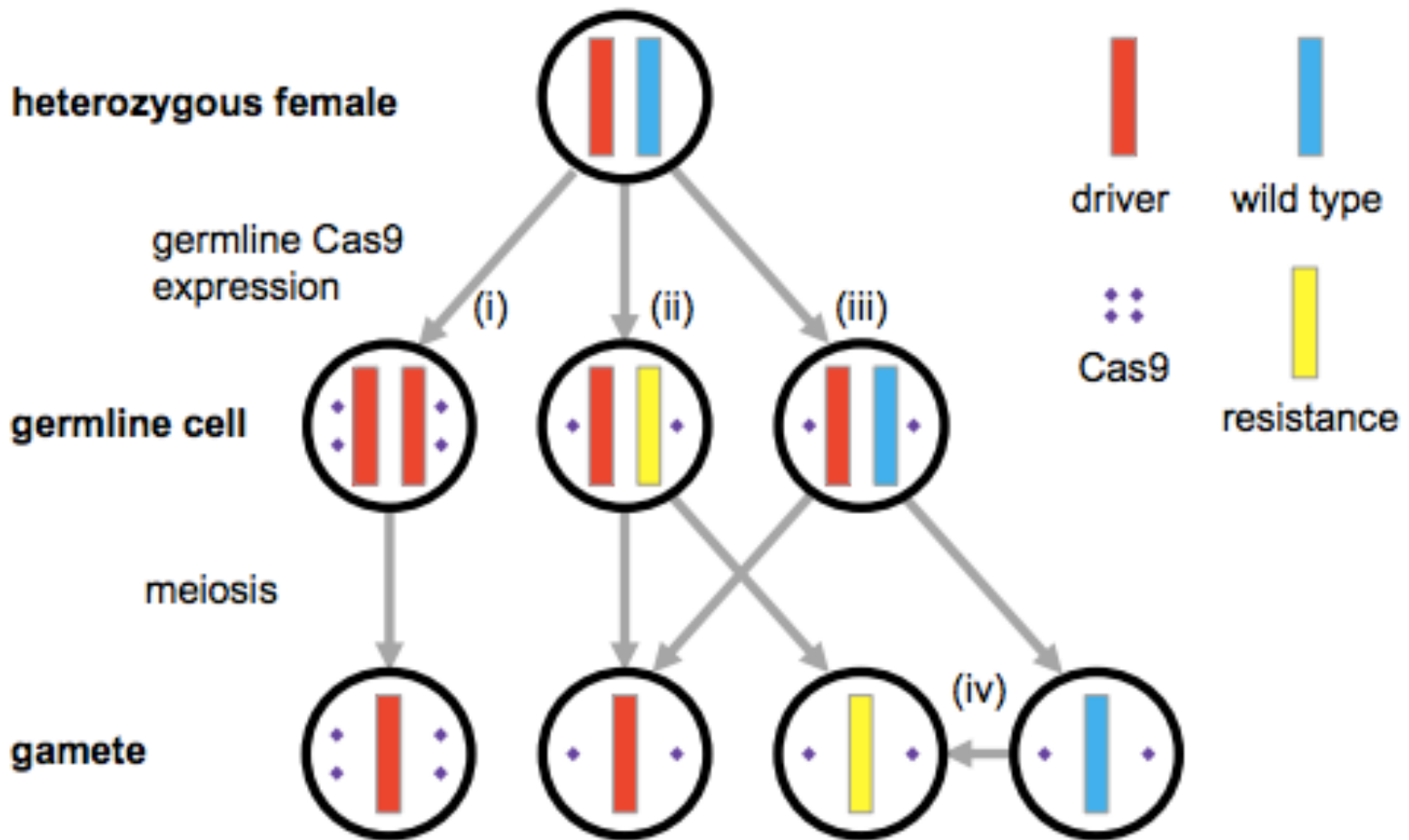


- **Public attitude surveys in Mali** suggest that people would like to see a **successful confined trial** before accepting a release:

*“I would have to see an example of modified mosquitoes reducing malaria in another village before I believe this claim”*

72-year-old man, Tienfala, Mali

## Q2. Can CRISPR-based gene drive be effective at disease control on a wide scale?



- Champer J, Reeves R, Oh SY, Liu C, Liu J *et al.* (2017) PLoS Genetics 13: e1006796

# Talk outline

## Overview

### Q1. Is it possible to perform a confined trial of a gene drive system?

- Threshold-dependent gene drive systems
- Inferring patterns of mosquito dispersal

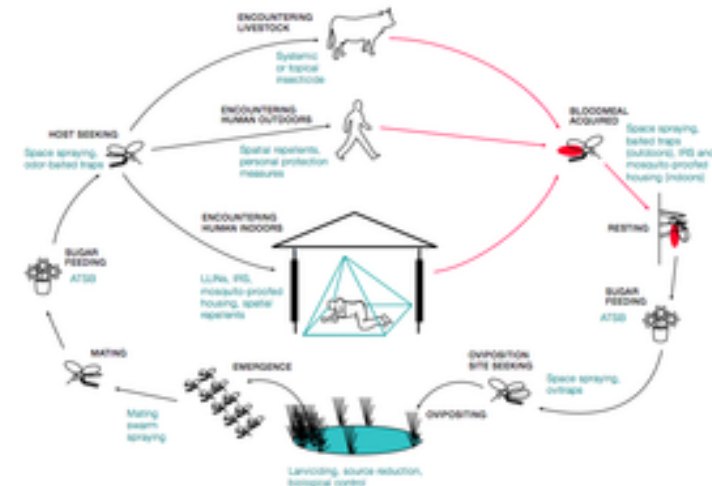
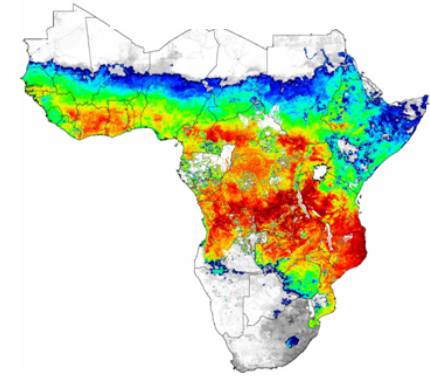
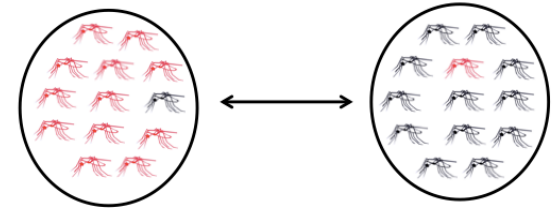
### Q2. Can CRISPR-based gene drive be effective at disease control on a wide scale?

- Resistant allele generation
- Possible solutions involving guide RNA multiplexing

### Q3. Which other novel vector control tools should we be prioritizing?

- Attractive toxic sugar baits (ATSB)
- Vector control optimization model (VCOM)

## Conclusion





# Q1. Is it possible to perform a confined trial of a gene drive system?

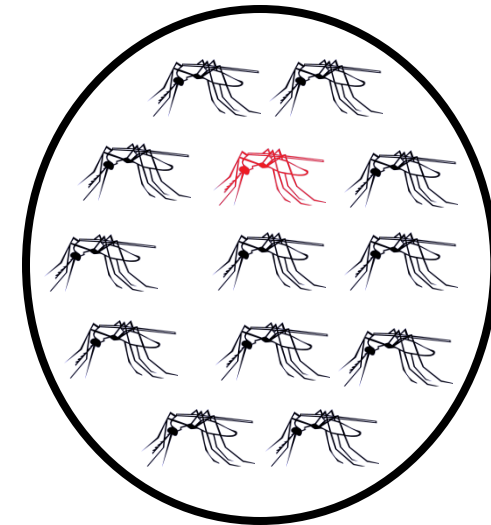
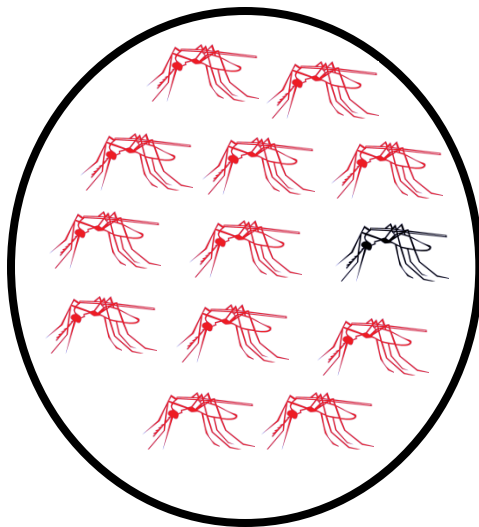


**Banambani, Mali**

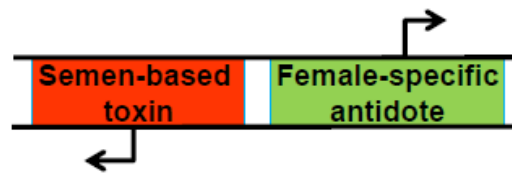
7 km  
~1% / gen



**Doneguebougou, Mali**



# Semele



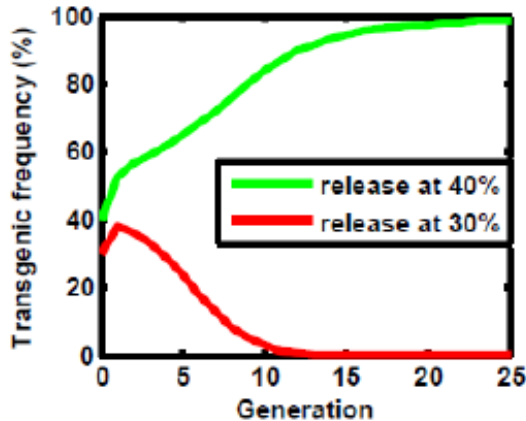
Transgenic males produce **toxic semen**

Transgenic females produce **antidote**

		Male			
		TT	Tt	tt	
Female	TT	TT <sub>1</sub>	TT <sub>2</sub>	Tt <sub>6</sub>	Tt <sub>5</sub>
	Tt	TT <sub>3</sub>	TT <sub>4</sub>	Tt <sub>8</sub>	Tt <sub>7</sub>
		Tt <sub>11</sub>	Tt <sub>8</sub>	tt <sub>14</sub>	tt <sub>13</sub>
	tt	Tt <sub>9</sub>	Tt <sub>10</sub>	tt <sub>12</sub>	tt

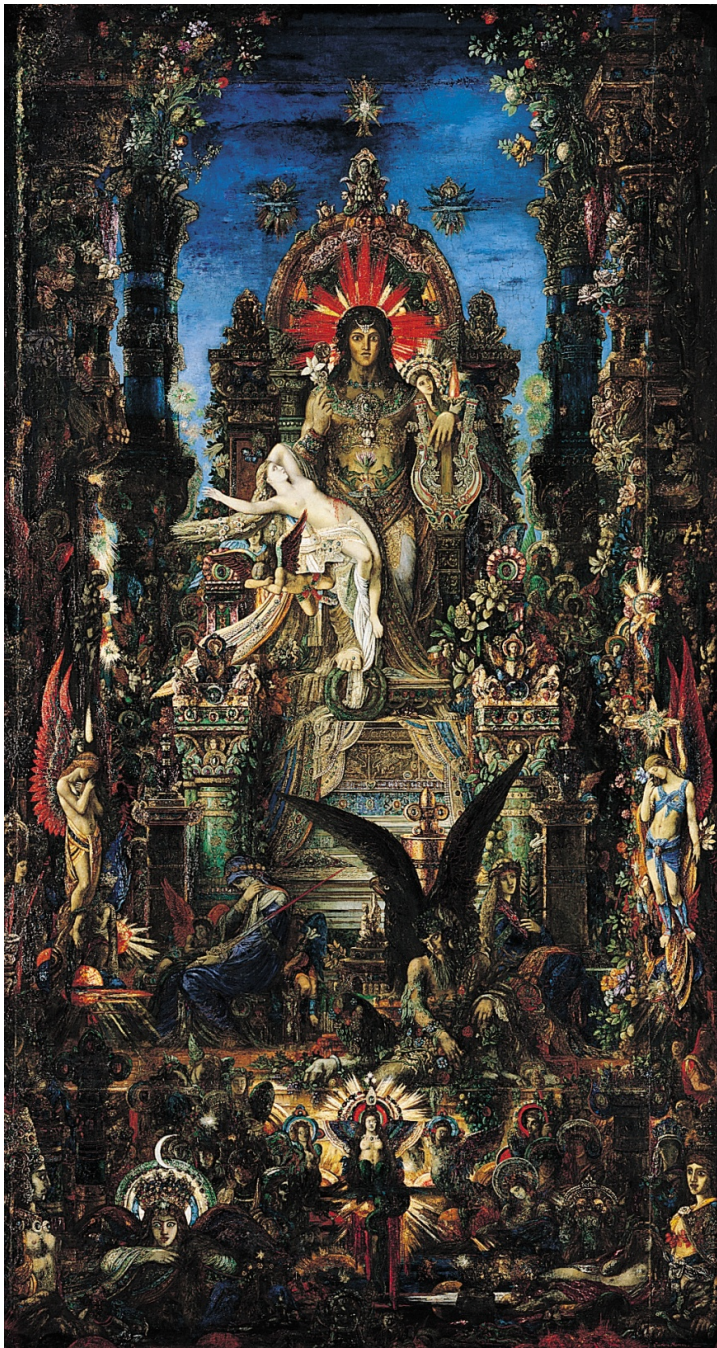
		Male			
		TT	Tt	tt	
Female	TT	TT <sub>1</sub>	TT <sub>2</sub>	Tt <sub>6</sub>	Tt <sub>5</sub>
	Tt	TT <sub>3</sub>	TT <sub>4</sub>	Tt <sub>8</sub>	Tt <sub>7</sub>
		Tt <sub>11</sub>	Tt <sub>8</sub>	tt <sub>14</sub>	tt <sub>13</sub>
	tt	Tt <sub>9</sub>	Tt <sub>10</sub>	tt <sub>12</sub>	tt

		Male			
		TT	Tt	tt	
Female	TT	TT <sub>1</sub>	TT <sub>2</sub>	Tt <sub>6</sub>	Tt <sub>5</sub>
	Tt	TT <sub>3</sub>	TT <sub>4</sub>	Tt <sub>8</sub>	Tt <sub>7</sub>
		Tt <sub>11</sub>	Tt <sub>8</sub>	tt <sub>14</sub>	tt <sub>13</sub>
	tt	<del>Tt<sub>9</sub></del>	<del>Tt<sub>10</sub></del>	<del>tt<sub>12</sub></del>	tt

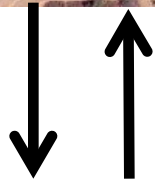


- A **release including females** results in **gene drive** (GM females are favored at high population frequencies).

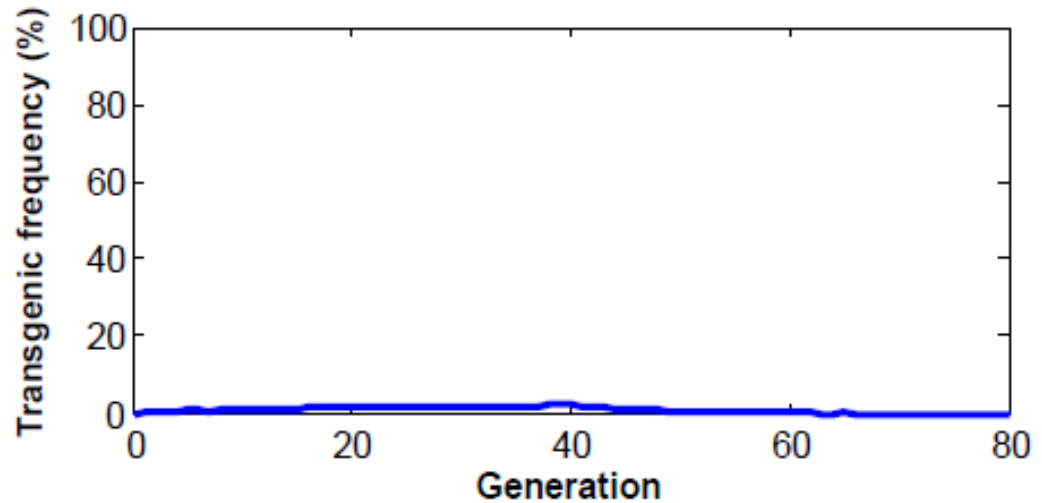
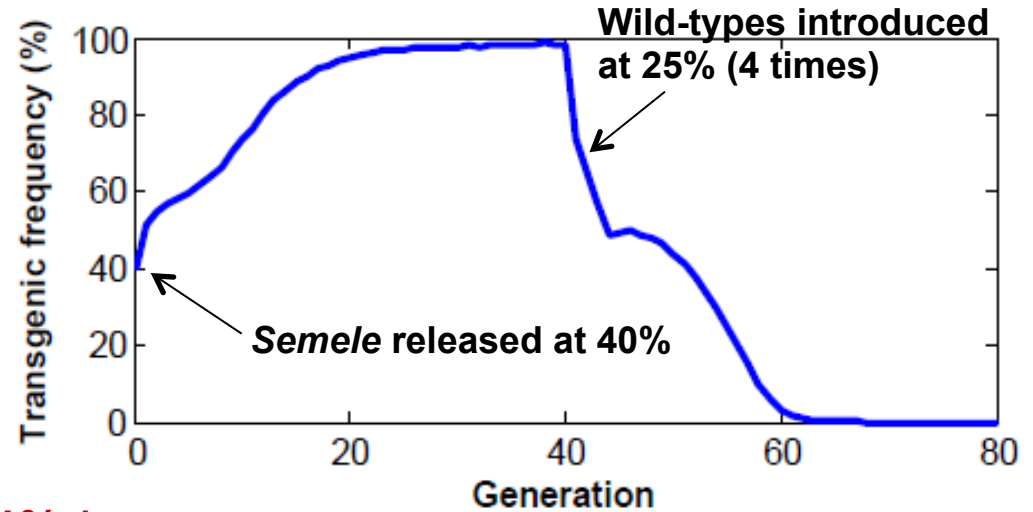
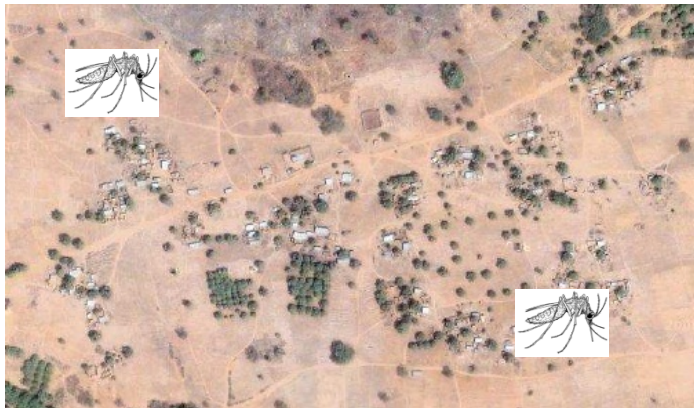
- **Release threshold = 36.4%:**



# Introduction of *Semele* is predicted to be confineable and reversible

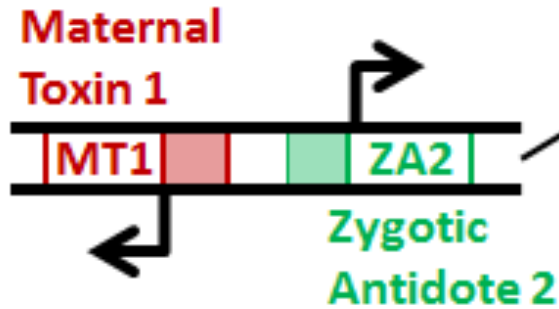


**Migration rate = 1% /gen**

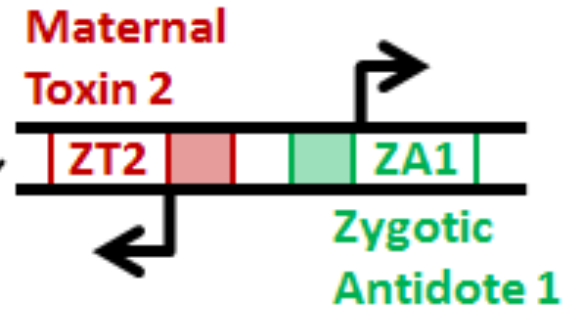


- Marshall JM, Pittman GW, Buchman A, Hay BA (2011) Genetics 187: 535-551.

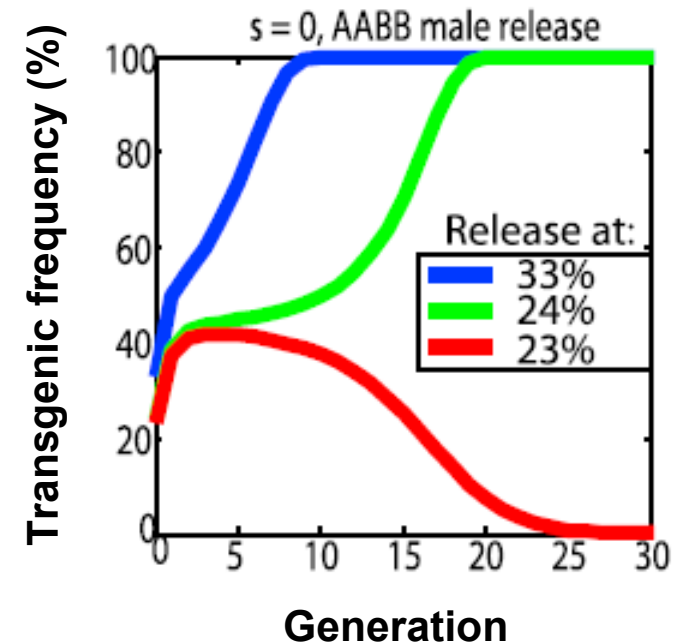
## Construct A:



## Construct B:



- Offspring of mothers having one or both constructs must inherit the opposite or both constructs to survive.
- This is more likely at higher population frequencies (>24%) leading to frequency-dependent drive.



# Inheritance pattern of UD<sup>MEL</sup>

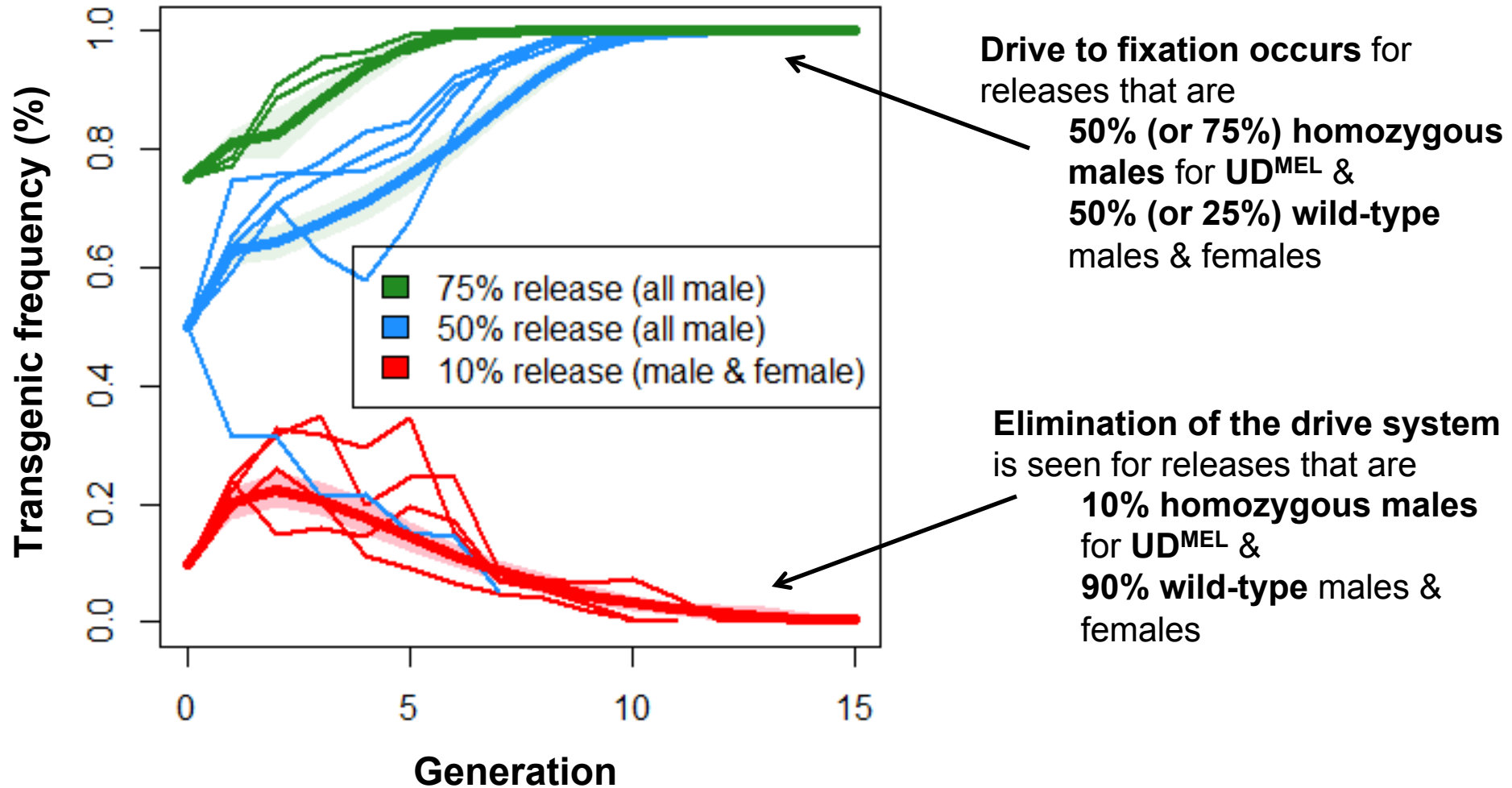
Female

		Female										
		A/A ; B/B	A/+ ; B/B	+/+ ; B/B	A/A ; B/+	A/A ; +/+	+/+ ; B/+	A/+ ; B/+	A/+ ; +/+	+/+ ; +/+		
Male	A/A ; B/B	A/A ; B/B	A/A ; B/B A/+ ; B/B		A/+ ; B/B	A/A ; B/B A/A ; B/+		A/A ; B/+	A/+ ; B/B A/+ ; B/+		A/A ; B/+ A/+ ; B/+	A/+ ; B/+
	A/+ ; B/B	A/A ; B/B	A/+ ; B/B		A/+ ; B/B	A/A ; B/B	A/A ; B/+	A/A ; B/+	A/+ ; B/B	A/+ ; B/+	A/A ; B/+	A/+ ; B/+
	+/+ ; B/B	A/+ ; B/B	A/A ; B/B	A/+ ; B/B	A/+ ; B/B	A/+ ; B/B	A/+ ; B/+	A/+ ; B/+	A/+ ; B/B	A/+ ; B/+	A/+ ; B/+	A/+ ; B/+
	A/A ; B/+	A/A ; B/B	A/A ; B/B	A/+ ; B/B	A/+ ; B/B	A/A ; B/B	A/A ; B/+	A/A ; B/+	A/+ ; B/B	A/+ ; B/+	A/A ; B/+	A/+ ; B/+
	A/A ; +/+	A/A ; B/+	A/A ; B/+		A/+ ; B/+	A/A ; B/+	A/A ; +/+	A/A ; +/+	A/+ ; B/+	A/+ ; +/+	A/A ; +/+	A/+ ; +/+
	+/+ ; B/+	A/+ ; B/B	A/+ ; B/B	A/+ ; B/B	A/+ ; B/B	A/+ ; B/+	A/+ ; B/+	A/+ ; B/+	A/+ ; B/B	A/+ ; B/+	A/+ ; B/+	A/+ ; +/+
	A/+ ; B/+	A/A ; B/B	A/+ ; B/B	A/A ; B/B	A/A ; B/+	A/+ ; B/B	A/+ ; B/+	A/A ; B/+	A/+ ; B/+	A/+ ; B/B	A/+ ; B/+	A/+ ; B/+
	A/+ ; +/+	A/A ; B/+	A/+ ; B/+		A/+ ; B/+	A/A ; B/+	A/A ; +/+	A/A ; +/+	A/+ ; B/+	A/+ ; +/+	A/A ; +/+	A/+ ; +/+
	+/+ ; +/+	A/+ ; B/+	A/+ ; B/+		A/+ ; B/+	A/+ ; B/+	A/+ ; +/+	A/+ ; +/+	A/+ ; B/+	A/+ ; +/+	A/+ ; +/+	A/+ ; +/+

2 Locus UD<sup>MEL</sup> 81 dihybrid punnet Square

- Akbari OS, Matzen KD, Marshall JM, Huang H *et al.* (2013) Current Biology 23: 671-677

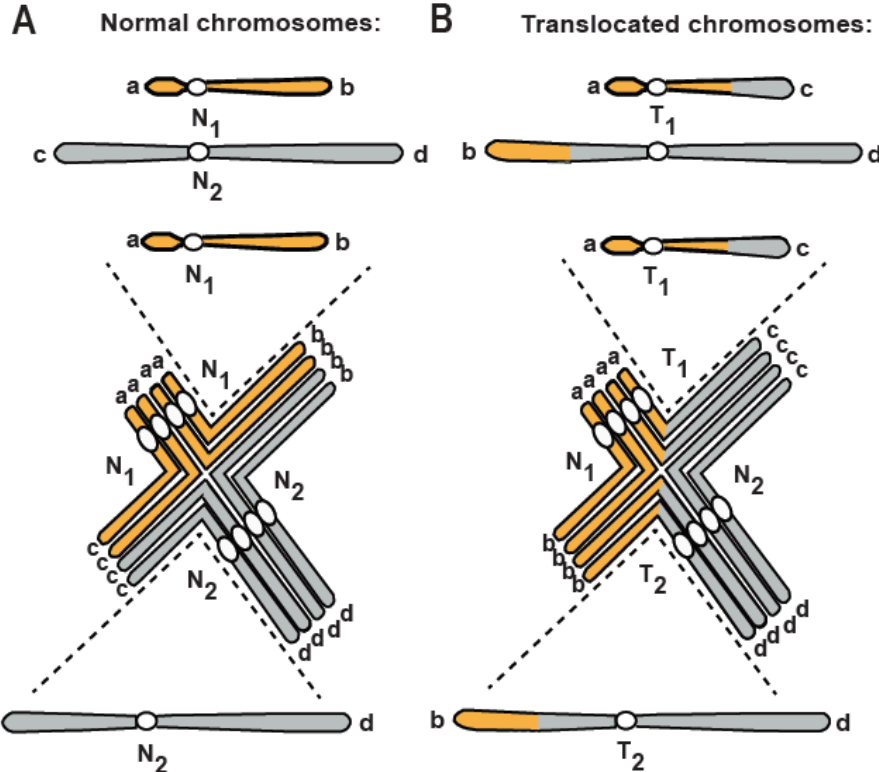
# UD<sup>MEL</sup> drive experiments agree with mathematical predictions & display threshold behavior



- Akbari OS, Matzen KD, Marshall JM, Huang H *et al.* (2013) *Current Biology* 23: 671-677

# Translocations also display threshold dynamics

Figure 1

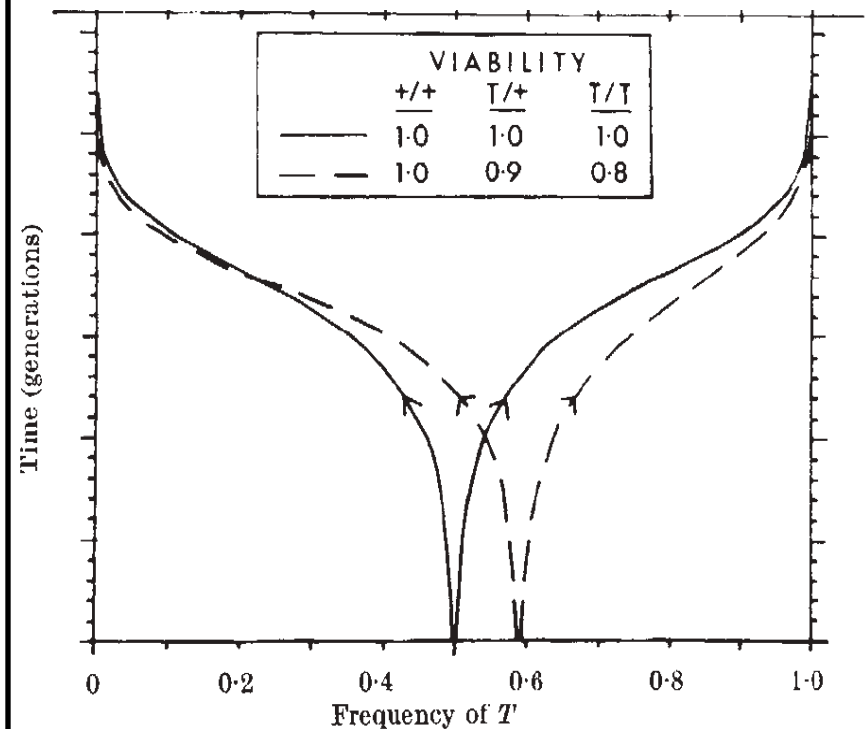


**C**

	$T_1N_2$	$N_1T_2$	$T_1T_2$	$N_1N_2$
$T_1N_2$	<del><math>T_1T_1N_2N_2</math></del>	$T_1N_1N_2T_2$	<del><math>T_1T_1N_2T_2</math></del>	<del><math>T_1N_1N_2N_2</math></del>
$N_1T_2$	$T_1N_1T_2N_2$	<del><math>N_1N_1T_2T_2</math></del>	<del><math>T_1N_1T_2T_2</math></del>	<del><math>N_1N_1T_2N_2</math></del>
$T_1T_2$	<del><math>T_1T_1T_2N_2</math></del>	$T_1N_1T_2T_2$	$T_1T_1T_2T_2$	$T_1N_1T_2N_2$
$N_1N_2$	<del><math>T_1N_1N_2N_2</math></del>	<del><math>N_1N_1N_2T_2</math></del>	$T_1N_1T_2N_2$	$N_1N_1N_2N_2$

NATURE, VOL. 218, APRIL 27, 1968

## Possible Use of Translocations to fix Desirable Genes in Insect Pest Populations

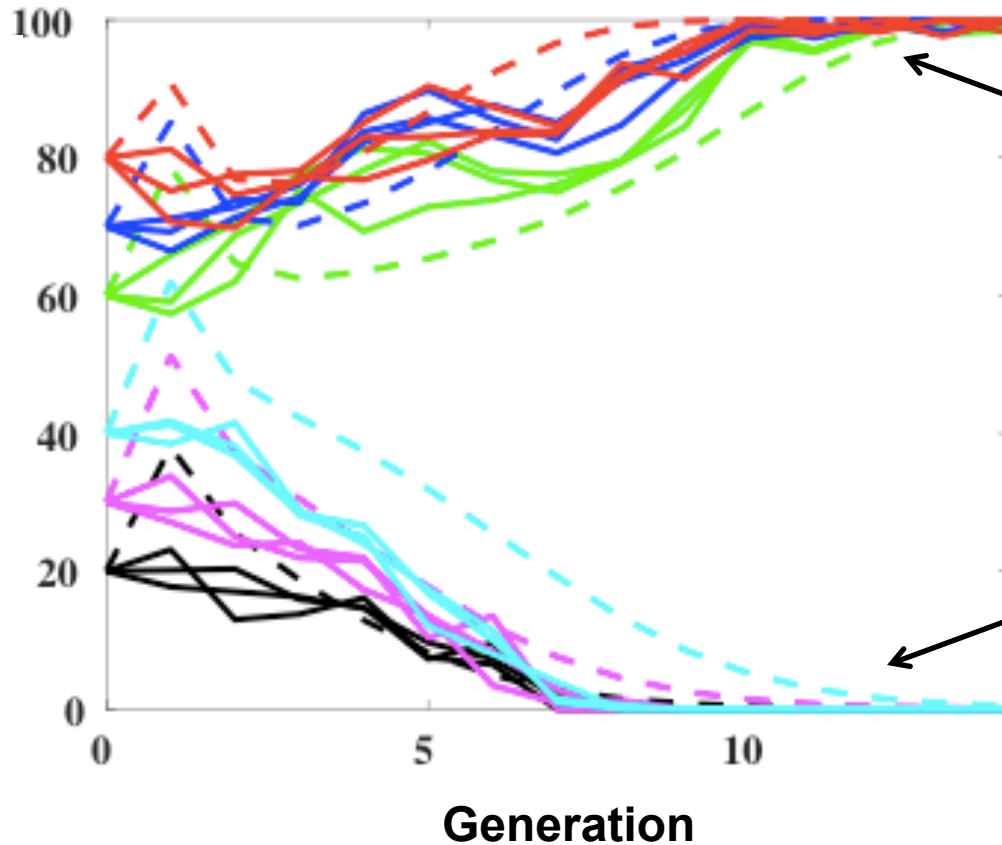


- Curtis CF (1968) Nature 218: 368-369
- Buchman A, Ivy T, Marshall JM, Akbari OS, Hay BA (2016) <http://dx.doi.org/10.1101/088393>



# Translocation drive experiments agree with model predictions & display threshold behavior

Frequency translocation bearing (%)



Drive to fixation occurs for releases that are **60-80% homozygous males & females** for the translocation

Elimination is seen for releases that are **20-40% homozygous males & females** for the translocation

(Translocation homozygotes are initially very unfit; but rapidly increase in fitness in a couple of generations)

- Buchman A, Ivy T, Marshall JM, Akbari OS, Hay BA (2016) <http://dx.doi.org/10.1101/088393>

# DARPA & “safe” gene drives

EXPLORE BY TAG



DEFENSE ADVANCED  
RESEARCH PROJECTS AGENCY

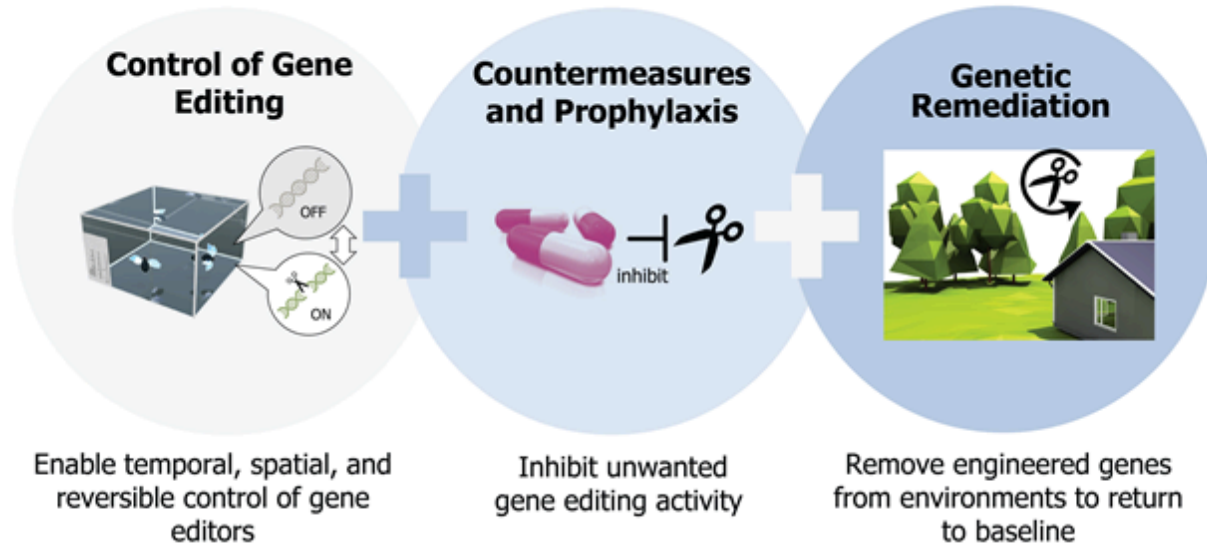
ABOUT US / OUR RESEARCH / NEWS / EVENTS / WORK WITH US /

Defense Advanced Research Projects Agency > News And Events

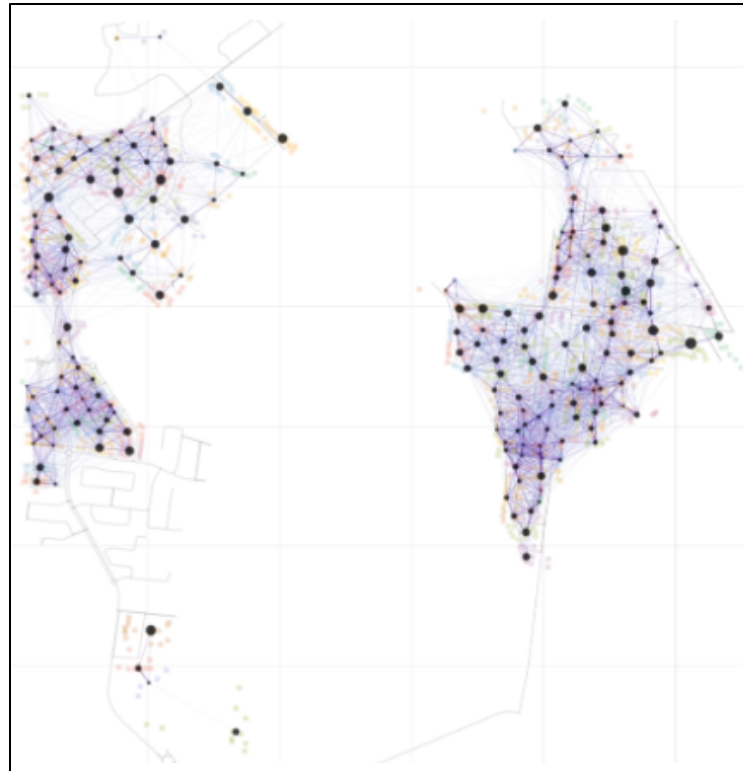
## Setting a Safe Course for Gene Editing Research

*Safe Genes program aims to build a biosafety and biosecurity toolkit to reduce potential risks and encourage innovation in the field of genome editing*

OUTREACH@DARPA.MIL  
9/7/2016



# MGDrivE modeling framework (Mosquito Gene Drive Explorer)



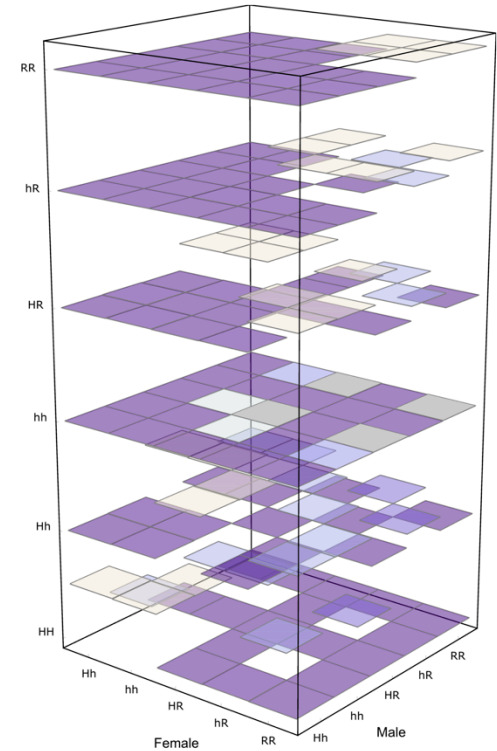
Spatial Setting

+



Mosquitoes

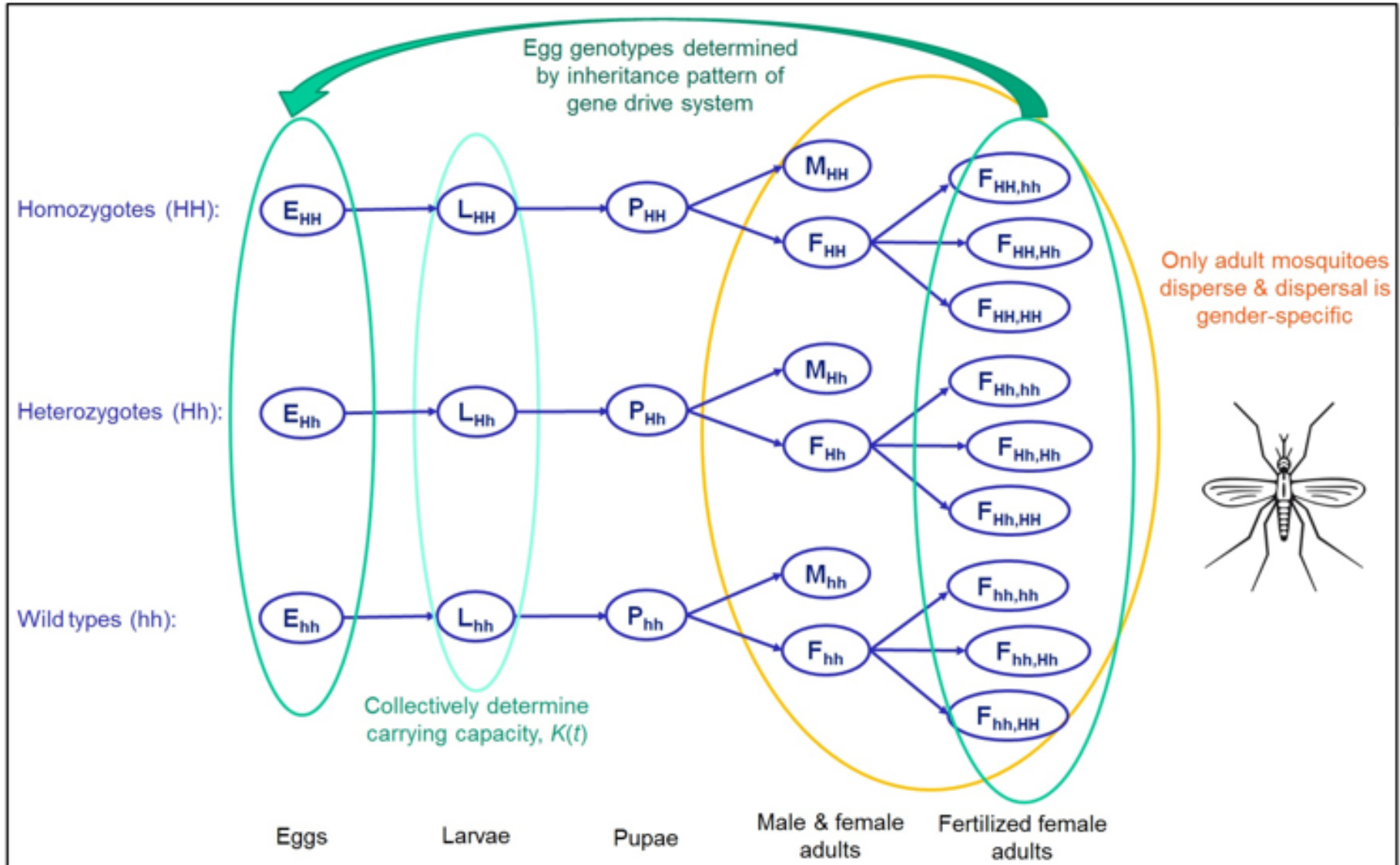
+



Genetic Inheritance

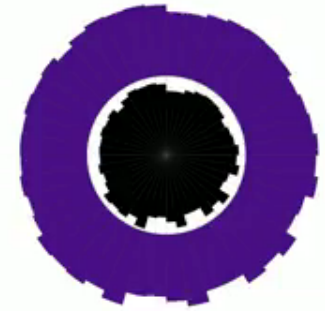
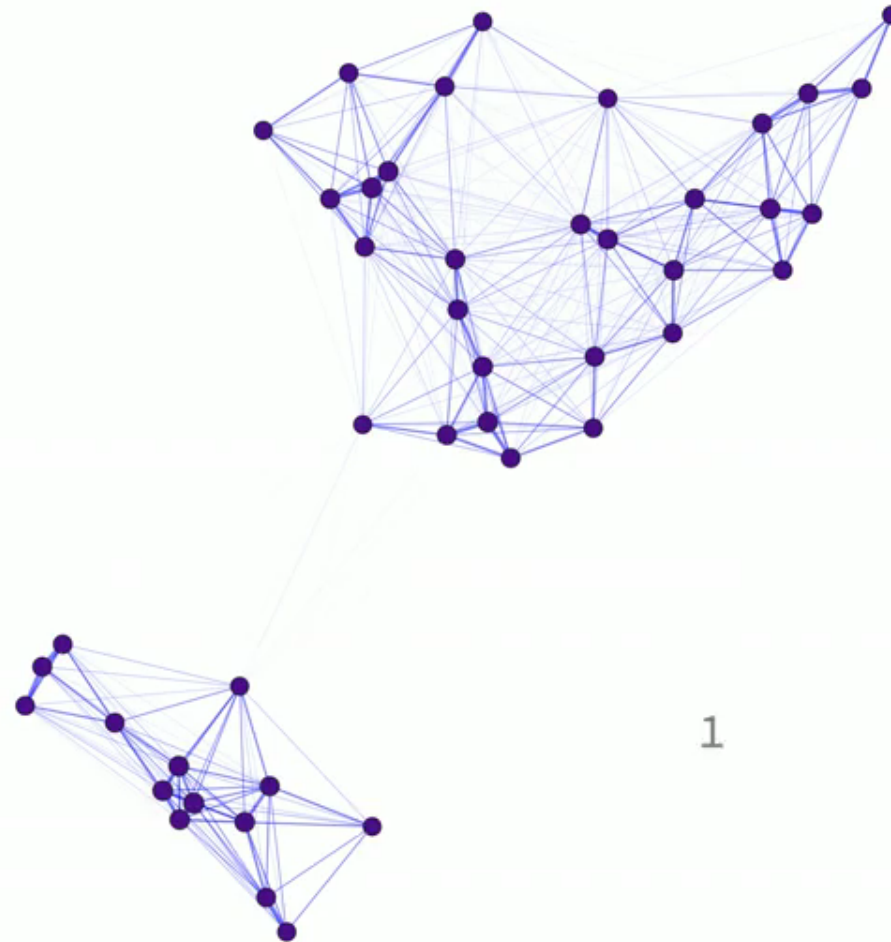
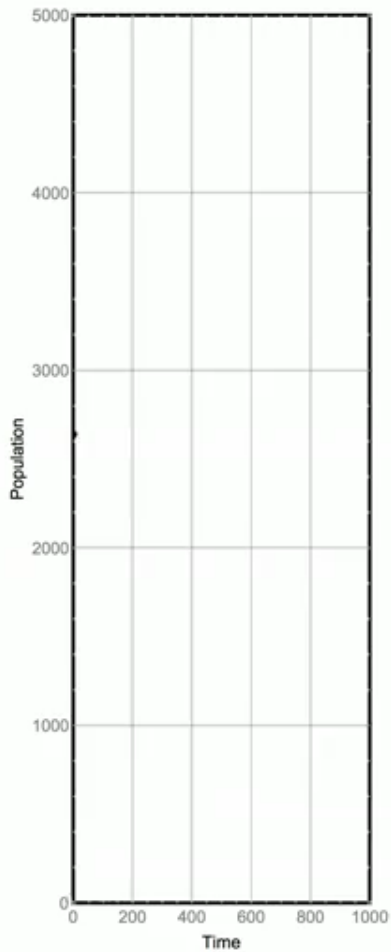
- Sanchez HM, Wu SL, Bennett J, Marshall JM (In preparation)

# MGDrivE: Mosquito ecology module

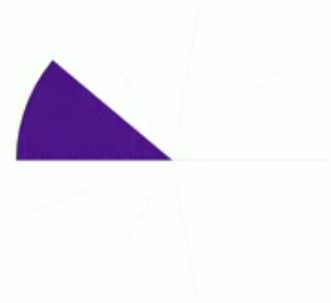


- Sanchez HM, Wu SL, Bennett J, Marshall JM (In preparation)

# MGDrivE: Translocations with remediation

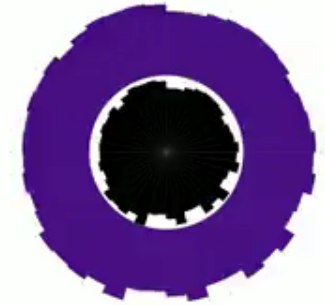
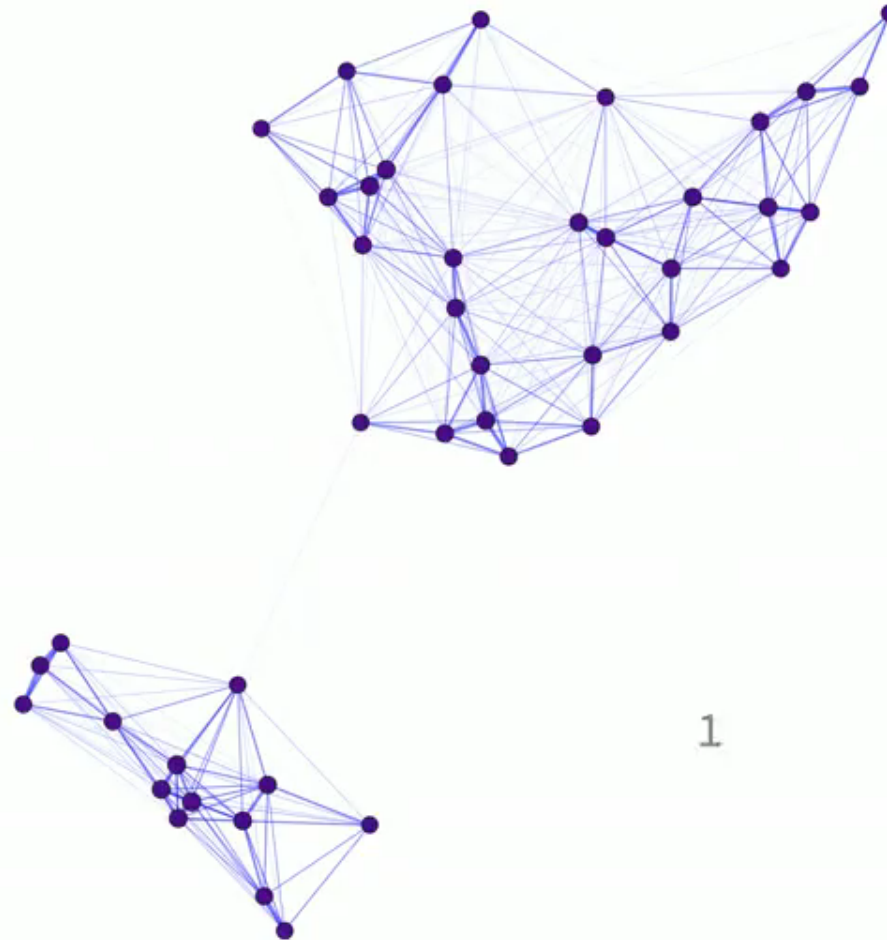
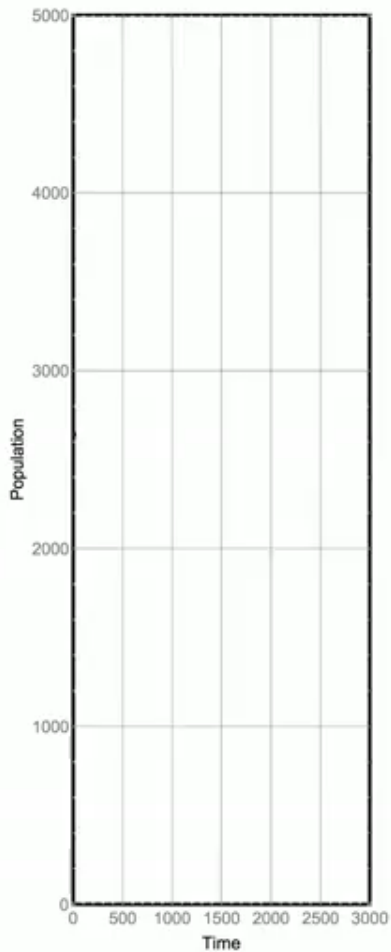


AABB	2640
AABb	0
AAbb	0
AaBB	0
AaBb	0
Aabb	0
aaBB	0
aaBb	0
aabb	0
Total	2640



- Sanchez HM, Wu SL, Bennett J, Marshall JM (In preparation)

# MGDrivE: UD<sup>MEL</sup> without remediation



AABB	0
AABb	0
AAbb	0
AaBB	0
AaBb	0
Aabb	0
aaBB	0
aaBb	0
aabb	2640
Total	2640

1

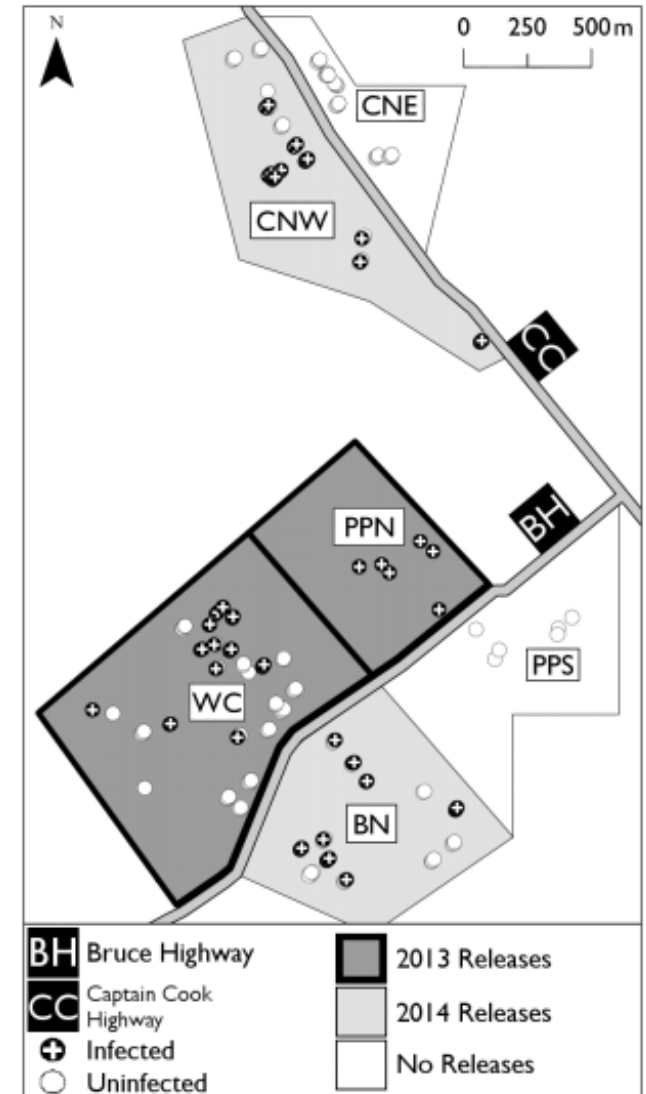
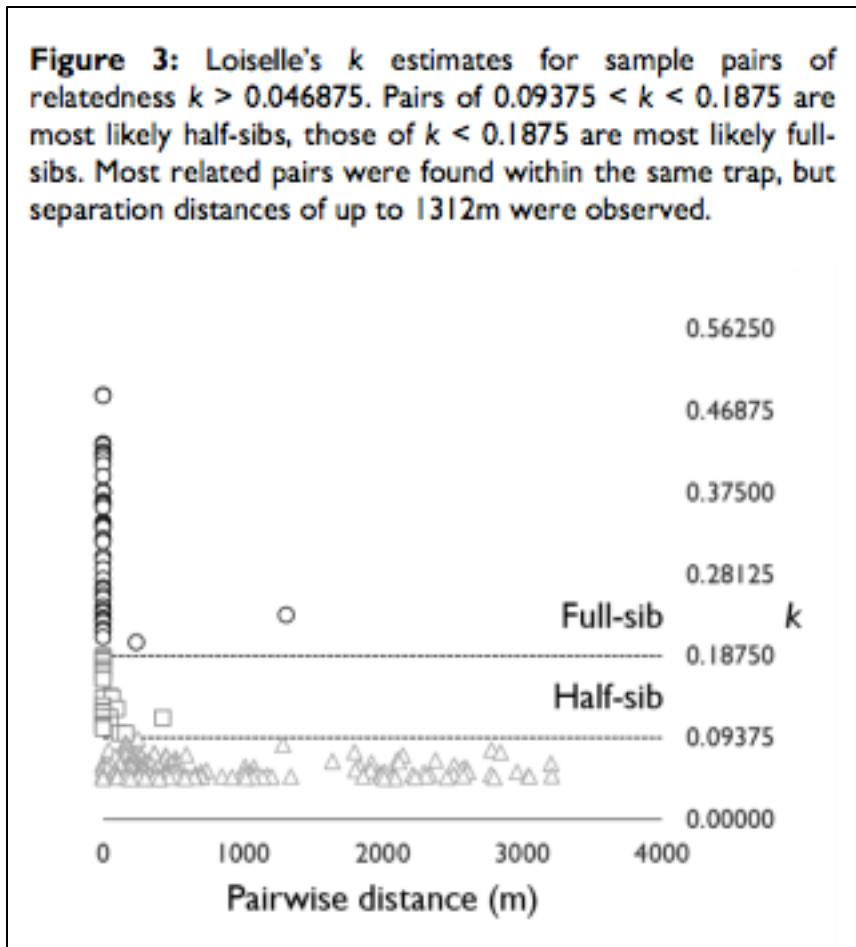


- Sanchez HM, Wu SL, Bennett J, Marshall JM (In preparation)

# Fine-scale landscape genomics helps explain the slow spread of *Wolbachia* through the *Aedes aegypti* population in Cairns, Australia

Thomas L Schmidt, Igor Filipovic, Ary A Hoffmann, Gordana Rasic

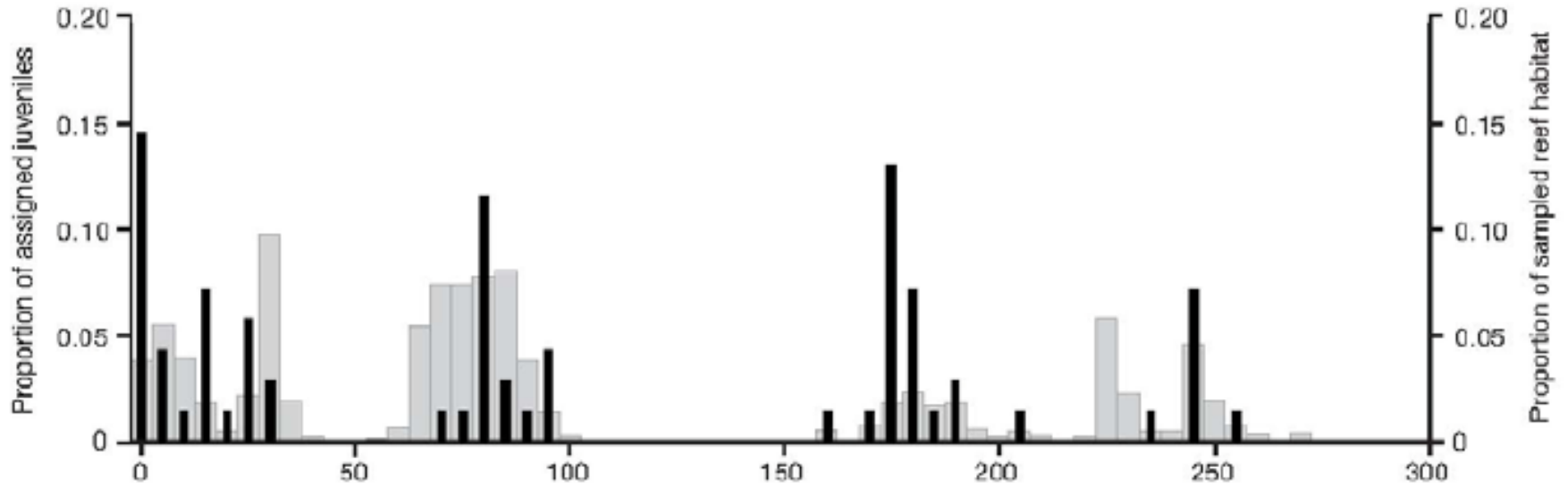
**Figure 3:** Loiselle's  $k$  estimates for sample pairs of relatedness  $k > 0.046875$ . Pairs of  $0.09375 < k < 0.1875$  are most likely half-sibs, those of  $k < 0.1875$  are most likely full-sibs. Most related pairs were found within the same trap, but separation distances of up to 1312m were observed.



- Schmidt TL, Filipovic I, Hoffmann AA, Rasic G (2017) <http://dx.doi.org/10.1101/103598>

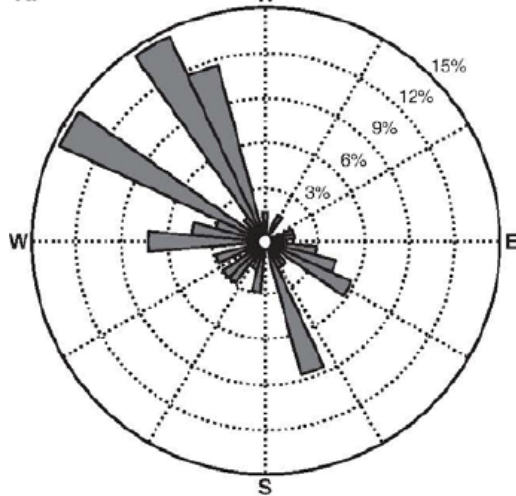
# Dispersal inferred from genetic parentage analyses

## A. *Plectropomus maculatus*



## *Plectropomus maculatus*

A.

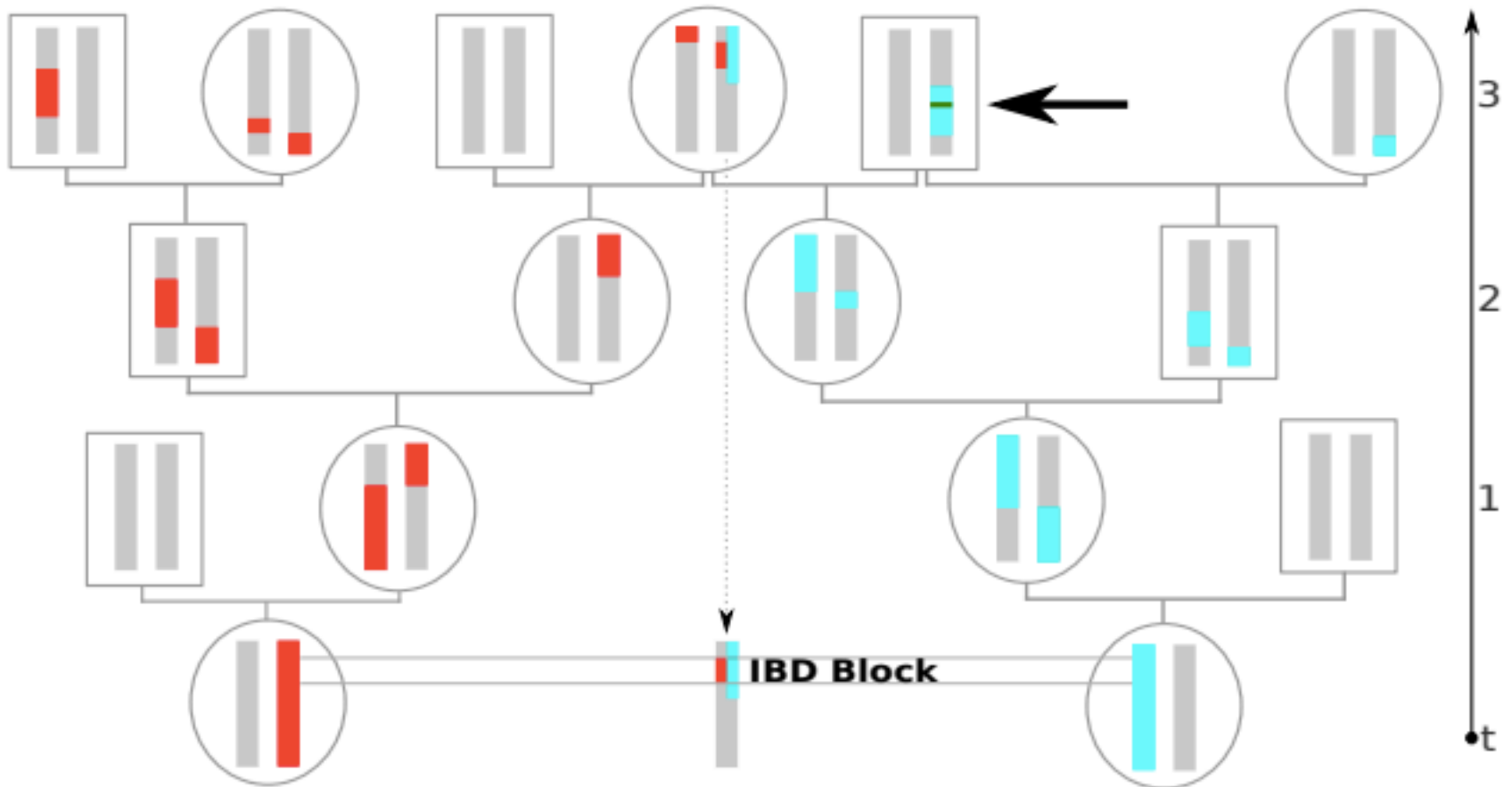


- Williamson DH, Harrison HB, Almany GR, Berumen ML *et al.* (2016) *Mol. Ecol.* 25L 6039-6054



# Inferring recent demography from isolation by distance of long shared sequence blocks

Harald Ringbauer<sup>\*,1</sup>, Graham Coop<sup>†</sup> and Nicholas H. Barton<sup>\*,1</sup>



- Ringbauer H, Coop G, Barton NH (2017) Genetics doi: 10.1534/genetics.116.196220.

# | UCI establishes Malaria Initiative to fight deadly disease in Africa

Acclaimed vector biologist Anthony James will lead multi-campus effort



## Team members



Ethan Bier (UCSD)  
UCSD site leader



Valentino Gantz (UCSD)  
Co-Investigator



John Marshall (UCB)  
Berkeley site leader



Yoosook Lee (UCD)  
Co-Investigator



Gregory Lanzaro (UCD)  
UCD site leader



Anthony Cornel (UCD)  
Co-Investigator



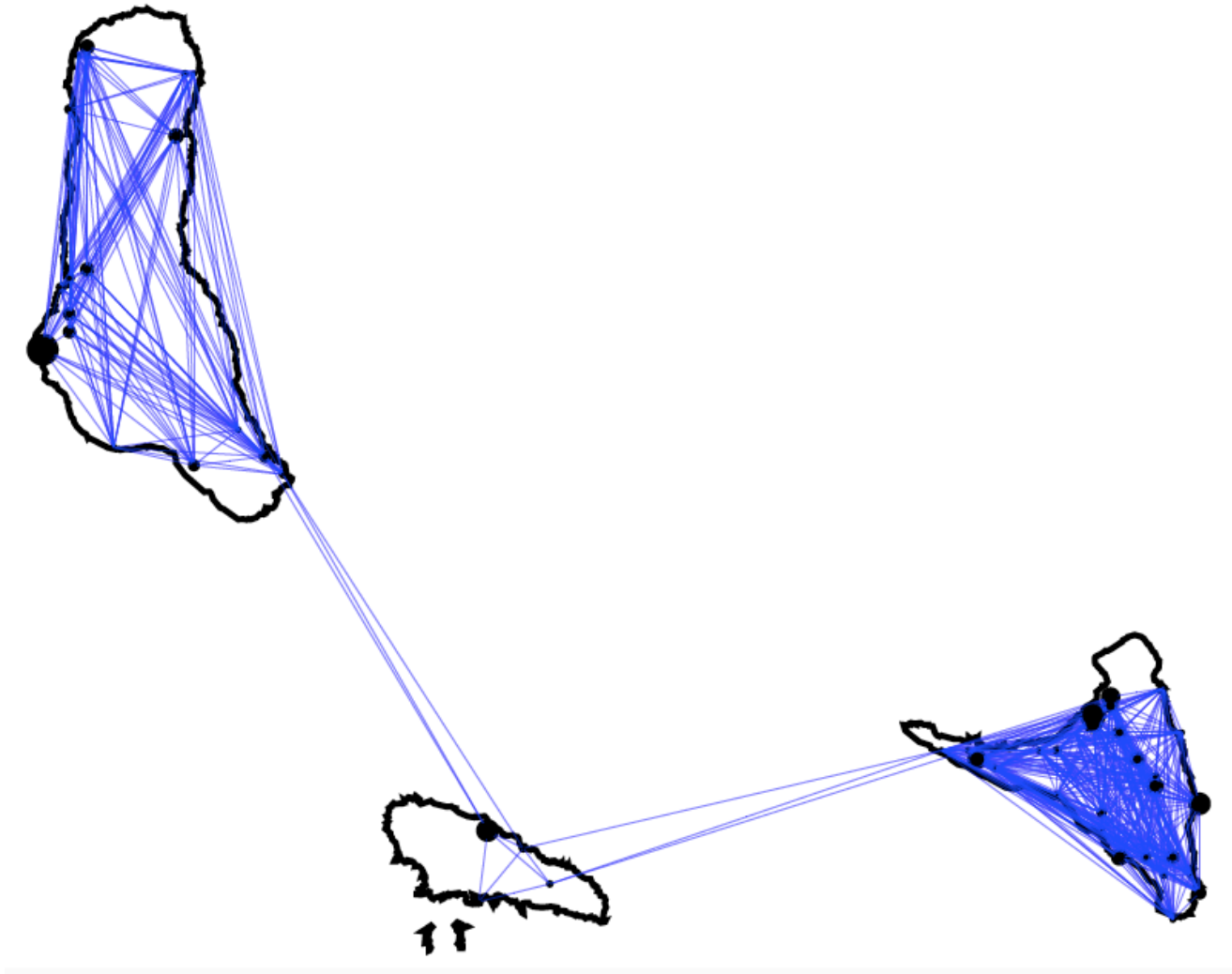
Ziad Haddad (UCLA)  
UCLA site leader



Sentelle Eubanks (UCI)  
Project Manager

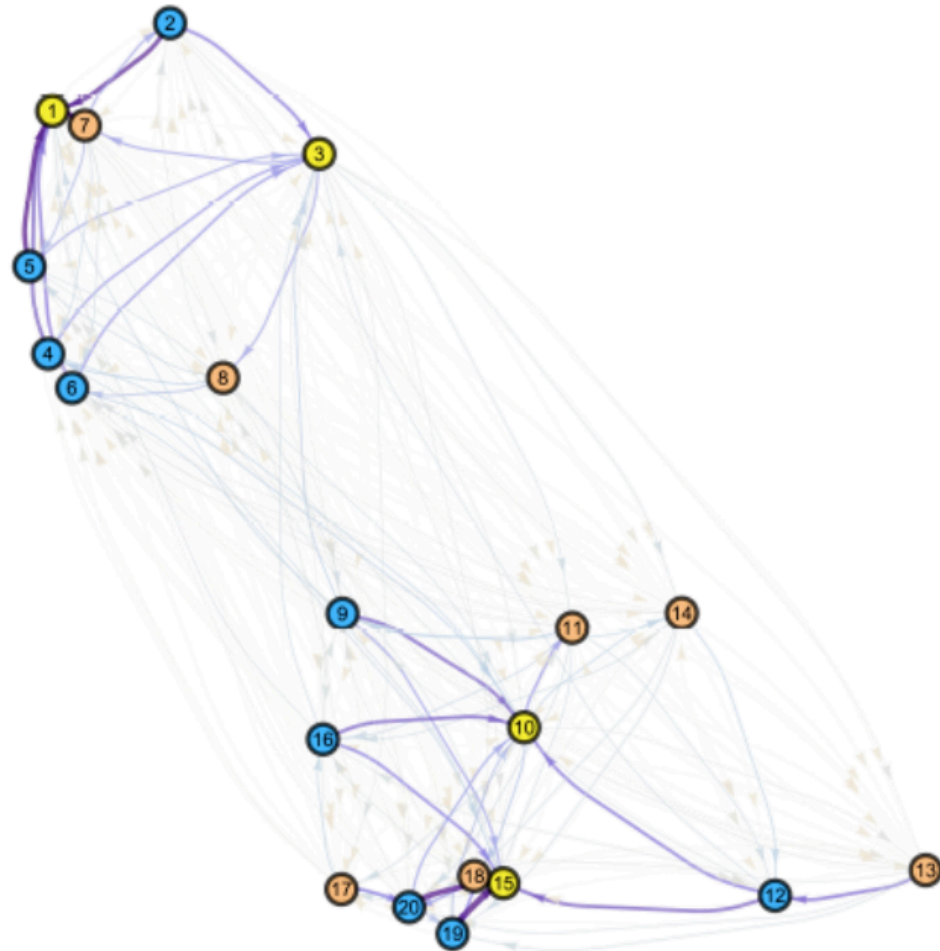
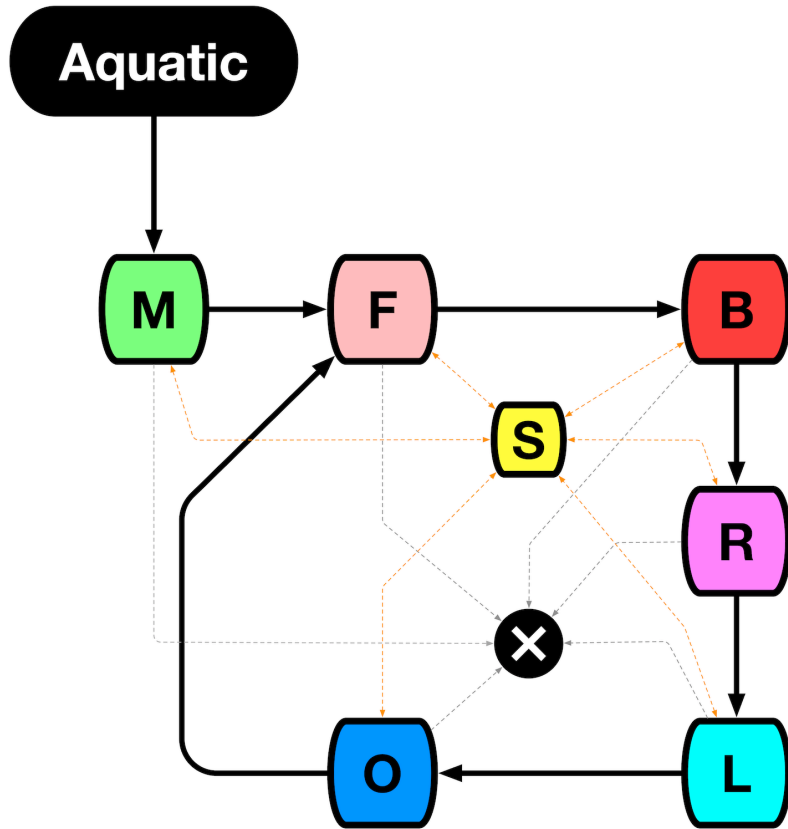
Population replacement strategy using homing-based gene drive.

# Application of MGDriVE to the Comoros Islands

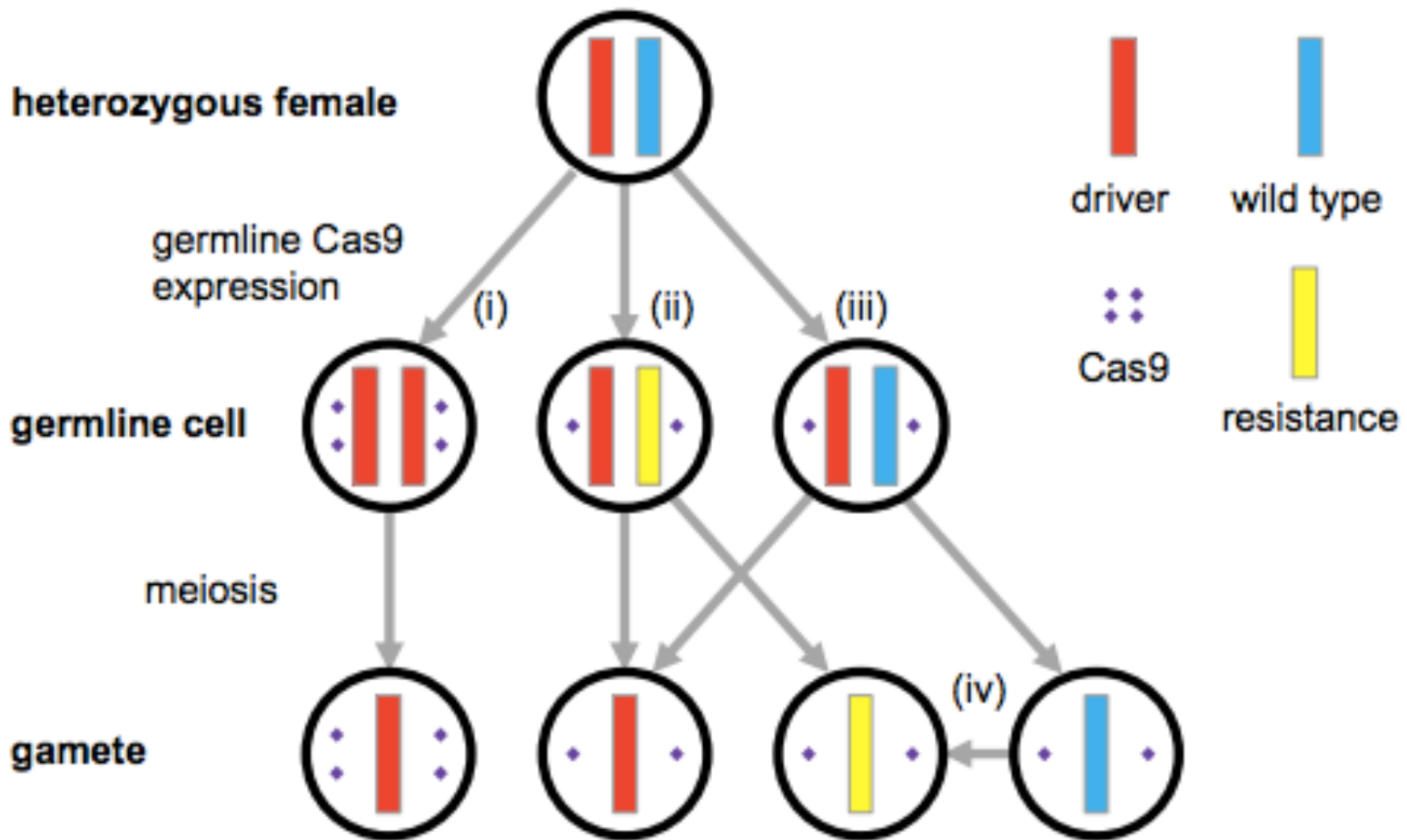


- Sanchez HM, Wu SL, Bennett J, Marshall JM (In preparation)

# Understanding potential fine-scale population structure using MASH modeling framework



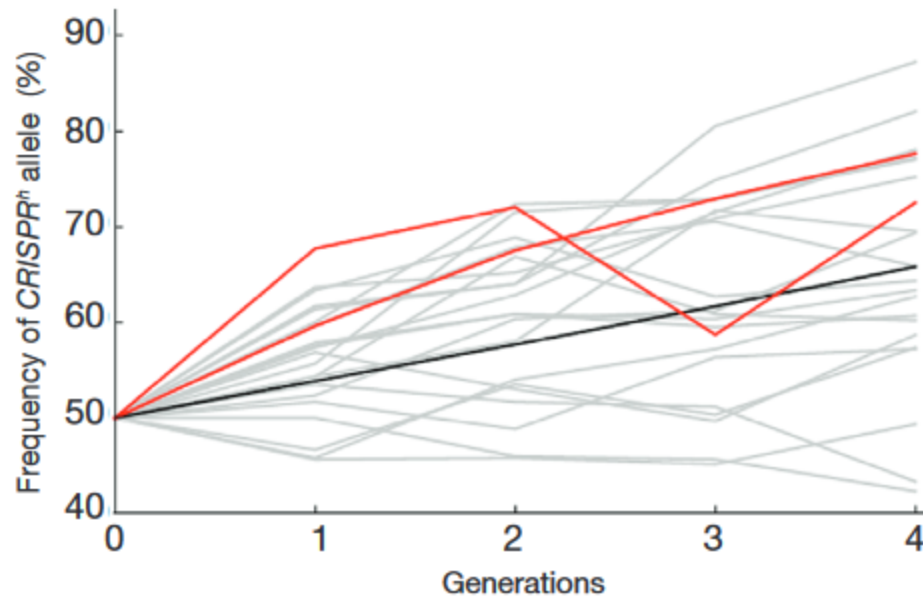
## Q2. Can CRISPR-based gene drive be effective at disease control on a wide scale?



- Champer J, Reeves R, Oh SY, Liu C, Liu J *et al.* (2017) PLoS Genetics 13: e1006796

# LETTERS

A CRISPR-Cas9 gene drive system targeting female reproduction in the malaria mosquito vector *Anopheles gambiae*



- Homing rate = 98%
- Non-cleavage rate = 1%
- Resistant allele generation rate = 0.13% (in-frame indels)
- Fertility of heterozygous females reduced by 90.7%

	Sequenced	Wild-type	Indel (independent)	Incomplete homing (independent)
AGAP011377	14	10	4 (3)	0
AGAP005958	13	5	8 (2)	0
AGAP007280	5	2	1 (1)	2 (1)

- Champer J, Reeves R, Oh SY, Liu C, Liu J *et al.* (2017) PLoS Genetics 13: e1006796

# Modeling error-prone homing-based gene drive

## Male

## Female

	HH	Hh	HR	hh	hR	RR
HH	1 HH	$\frac{(1+e)}{2}$ HH $\frac{(1-e-p)}{2}$ Hh $\frac{p}{2}$ HR	$\frac{1}{2}$ HH $\frac{1}{2}$ HR	(1) Hh	$\frac{1}{2}$ Hh $\frac{1}{2}$ HR	(1) HR
Hh	$\frac{(1+e)2p}{4}$ HH $\frac{(1-e-p)}{2}$ Hh $\frac{p}{2}$ HR	Cross A	Cross B	$\frac{(1+e)}{2}$ Hh $\frac{(1-e-p)}{2}$ hh $\frac{p}{2}$ hR	Cross D	$\frac{(1+e)}{2}$ HR $\frac{(1-e-p)}{2}$ hR $\frac{p}{2}$ RR
HR	$\frac{1}{2}$ HH $\frac{1}{2}$ HR	Cross C	$\frac{1}{4}$ HH $\frac{1}{2}$ HR $\frac{1}{4}$ RR	$\frac{1}{2}$ Hh $\frac{1}{2}$ hR	$\frac{1}{4}$ Hh $\frac{1}{4}$ HR $\frac{1}{4}$ hR $\frac{1}{4}$ RR	$\frac{1}{2}$ HR $\frac{1}{2}$ RR
hh	(1) Hh	$\frac{(1+e)}{2}$ Hh $\frac{(1-e-p)}{2}$ hh $\frac{p}{2}$ HR	$\frac{1}{2}$ Hh $\frac{1}{2}$ hR	(1) hh	$\frac{1}{2}$ hR $\frac{1}{2}$ hh	(1) hR
hR	$\frac{1}{2}$ Hh $\frac{1}{2}$ HR	Cross E	$\frac{1}{4}$ Hh $\frac{1}{4}$ HR $\frac{1}{4}$ hR $\frac{1}{4}$ RR	$\frac{1}{2}$ hR $\frac{1}{2}$ hh	$\frac{1}{4}$ hh $\frac{1}{2}$ hR $\frac{1}{4}$ RR	$\frac{1}{2}$ hR $\frac{1}{2}$ RR
RR	(1) HR	$\frac{(1+e)}{2}$ HR $\frac{(1-e-p)}{2}$ hR $\frac{p}{2}$ RR	$\frac{1}{2}$ HR $\frac{1}{2}$ RR	(1) hR	$\frac{1}{2}$ hR $\frac{1}{2}$ RR	(1) RR

### Cross A

$\frac{((1+e)^2)}{4}$  HH       $\frac{((1-e-p)^2)}{4}$  hh  
 $\frac{((1+e)(1-e-p))}{2}$  Hh     $\frac{((1-e-p)p)}{2}$  hR  
 $\frac{((1+e)p)}{2}$  HR       $\frac{(p^2)}{4}$  RR

### Cross B/C

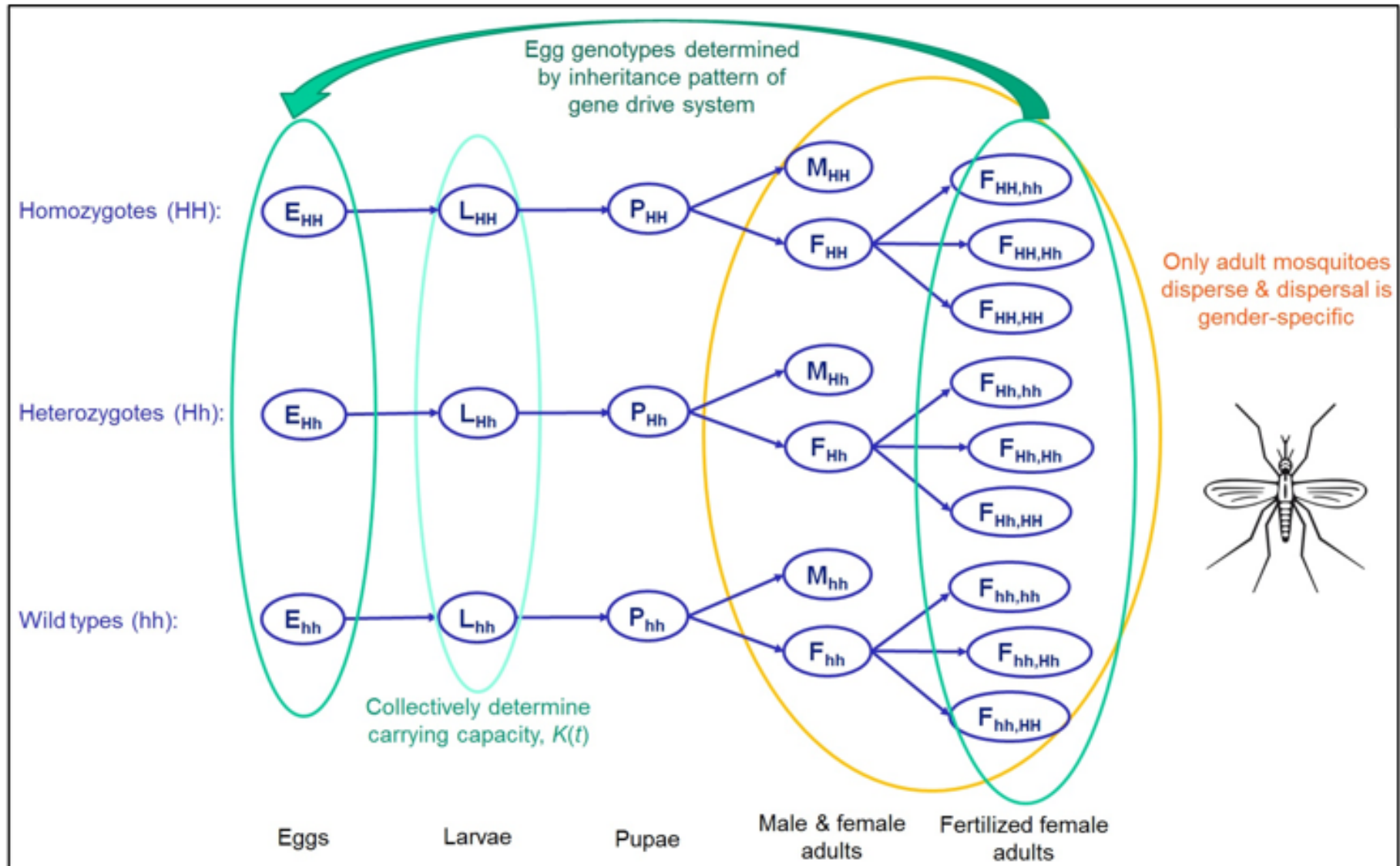
$\frac{(1+e)}{4}$  HH       $\frac{(1-e-p)}{4}$  hR  
 $\frac{(1-e-p)}{4}$  Hh       $\frac{(p)}{4}$  RR  
 $\frac{(1+e+p)}{4}$  HR

### Cross D/E

$\frac{(1+e)}{4}$  Hh       $\frac{(1-e-p)}{4}$  hh  
 $\frac{(1+e)}{4}$  HR       $\frac{(1-e)}{4}$  hR  
 $\frac{(p)}{4}$  RR

- Marshall JM, Buchman A, Sanchez HM *et al.* (2017) Nature Sci Rep 7: 3776

# MGDrivE: Mosquito ecology module



- Marshall JM, Buchman A, Sanchez HM *et al.* (2017) Nature Sci Rep 7: 3776



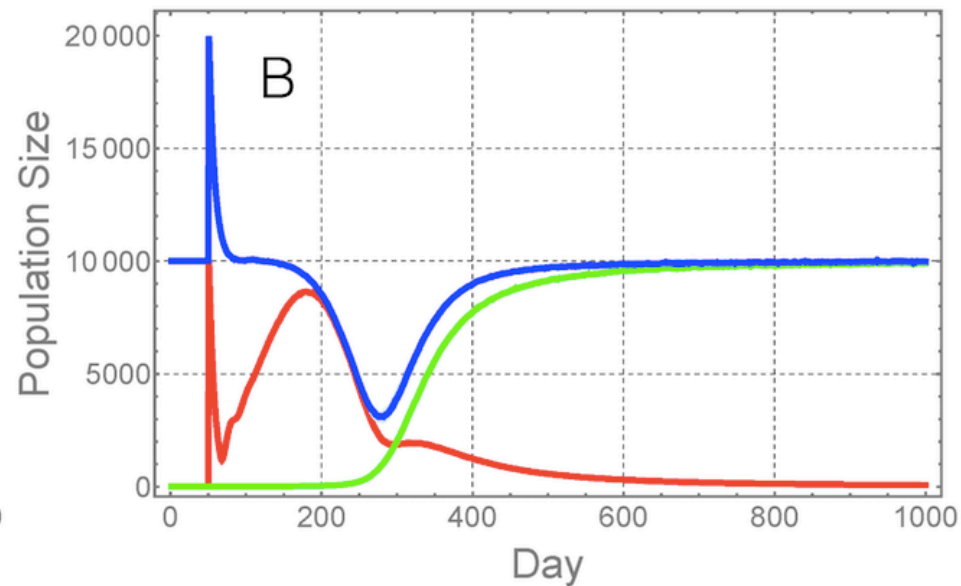
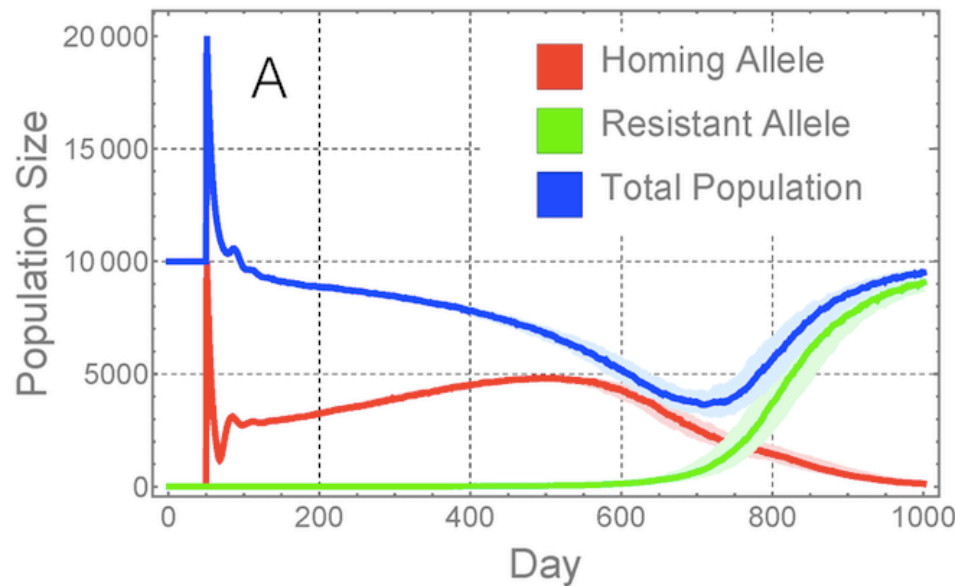
# Dynamics of current constructs

a) Hammond *et al.* (2016) construct:

- Homing rate = 98%
- NHEJ rate = 0.13%
- Fertility of heterozygotes reduced by 90.7%

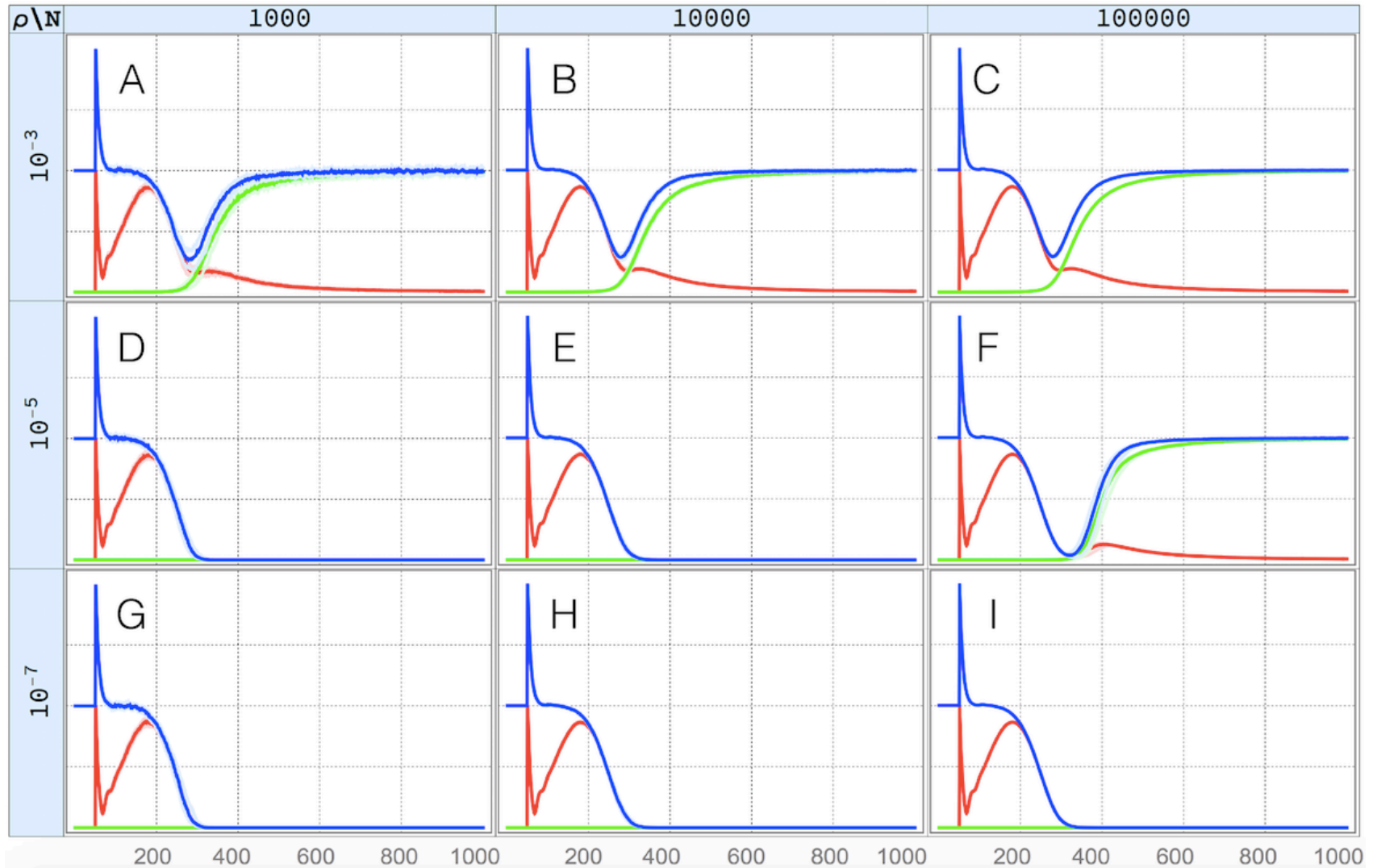
b) Hammond *et al.* (2016) construct:

- Homing rate = 98%
- NHEJ rate = 0.13%
- **Fertility of heterozygotes same as wild-type**



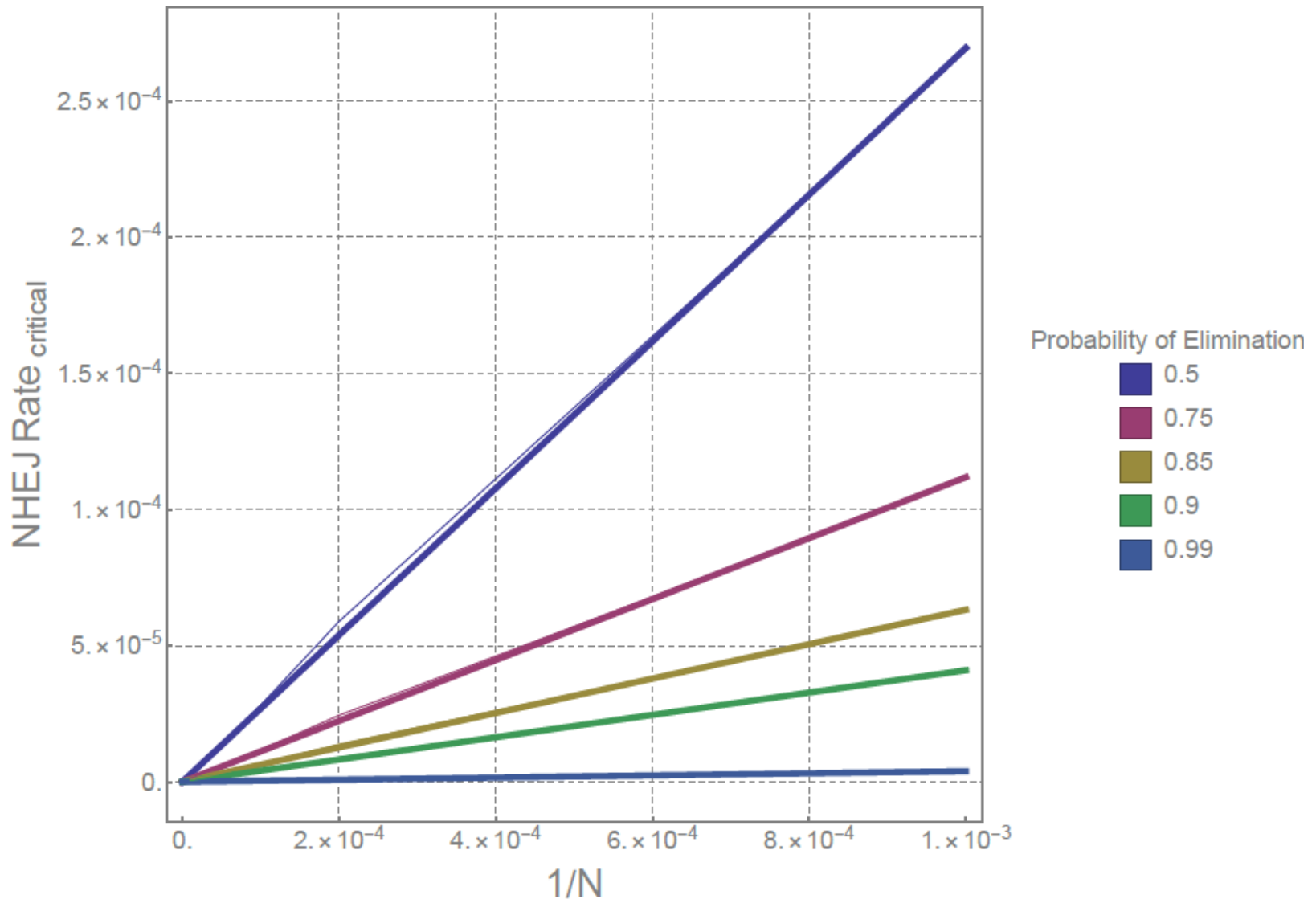
- Marshall JM, Buchman A, Sanchez HM *et al.* (2017) Nature Sci Rep 7: 3776

# As the resistance allele generation rate declines, the population size you can eliminate increases



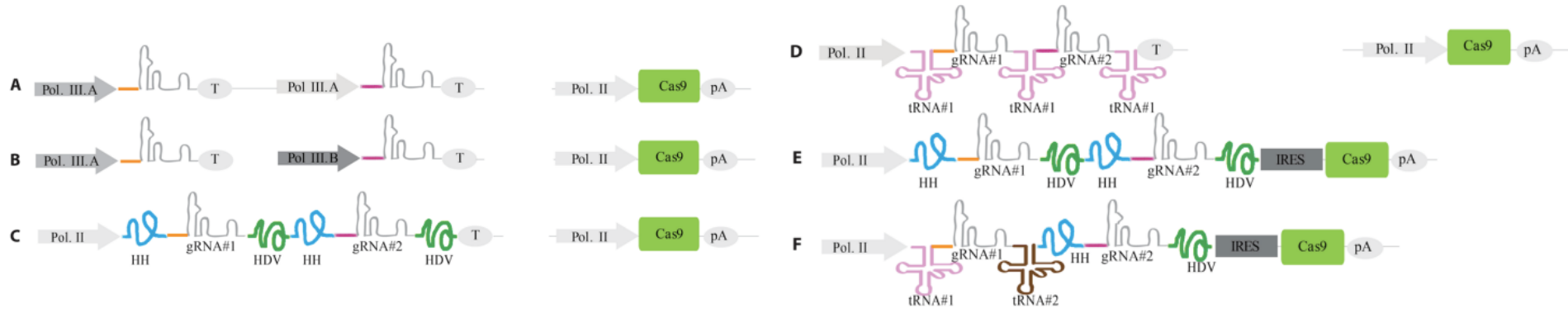
- Marshall JM, Buchman A, Sanchez HM *et al.* (2017) Nature Sci Rep 7: 3776

# Tolerable rates of resistant allele generation are inversely proportional to the population size



- Marshall JM, Buchman A, Sanchez HM *et al.* (2017) Nature Sci Rep 7: 3776

# Multiplexing gRNAs may provide part of the solution to enable elimination of large populations

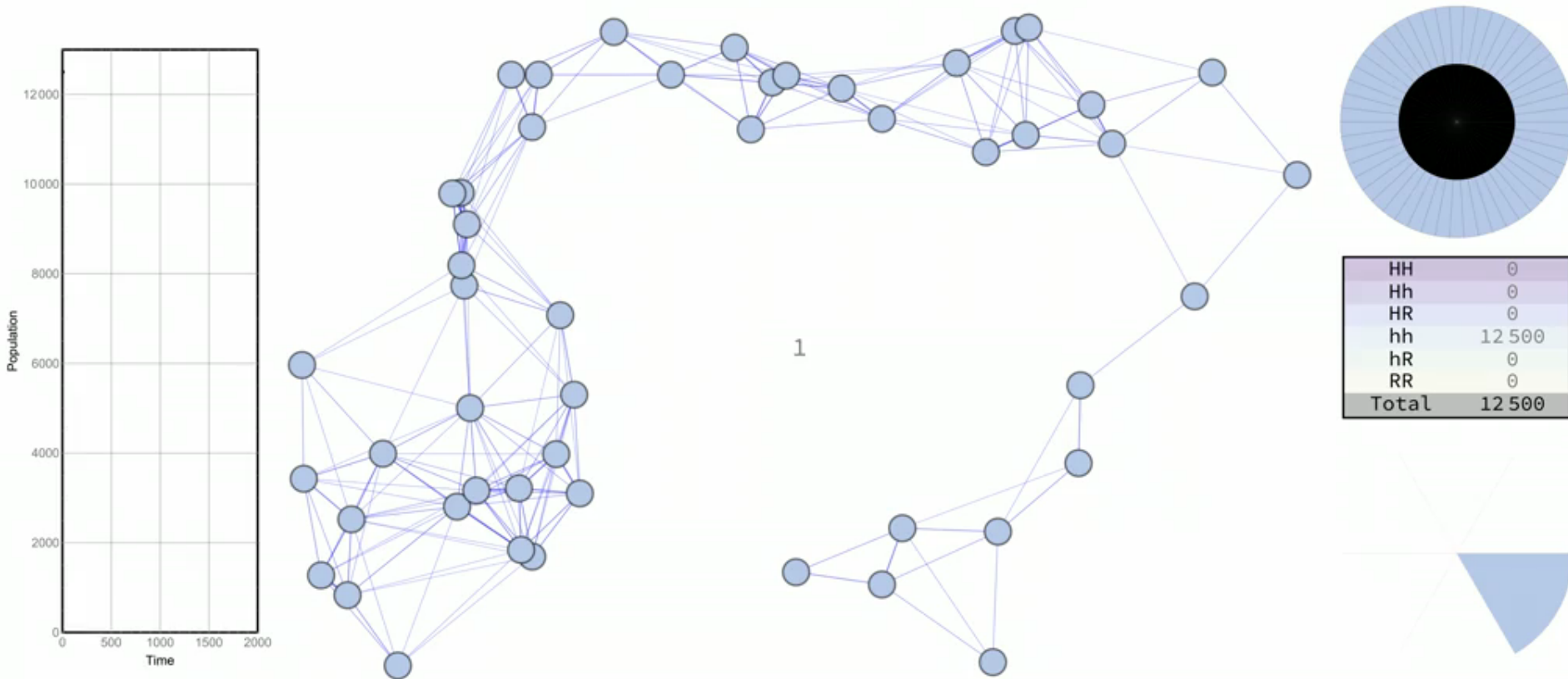


Multiplex number:	Resistance allele generation rate:	Population size capable of eliminating (90% of sims):
1	$1.3 \times 10^{-3}$	32
2	$1.7 \times 10^{-6}$	24 thousand
3	$2.2 \times 10^{-9}$	19 million
4	$2.9 \times 10^{-12}$	14 billion

- Marshall JM, Buchman A, Sanchez HM *et al.* (2017) Nature Sci Rep 7: 3776

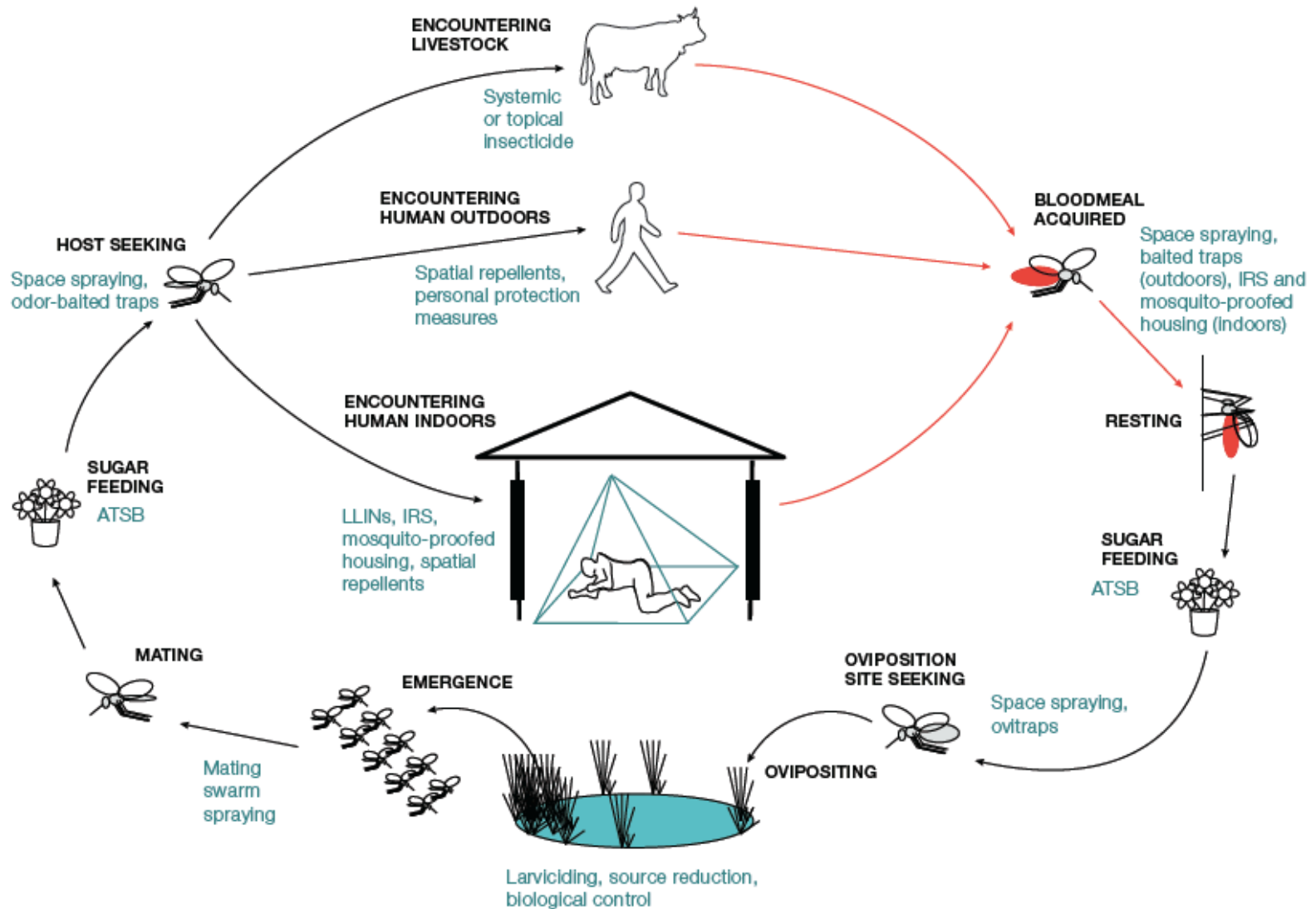
# MGDrive: Homing-based drive targeting a female fertility gene with resistance allele generation

## MGDrive



- Sanchez HM, Wu SL, Bennett J, Marshall JM (In preparation)

# Q3. Which other novel vector control tools should we be prioritizing?



- Kiware SS, Chitnis C, Tatarsky A, Wu SL, Sanchez HM *et al.* (2017) PLoS ONE 12: e0187680

# Attractive toxic sugar baits (ATSB)

- **Attract with:**

- **Fruity or flowery scent (bait)**



- **Then kill with:**

- **Sugar (feeding stimulant)**
- **Boric acid (oral toxin)**



- **Tested extensively in Israel on:**

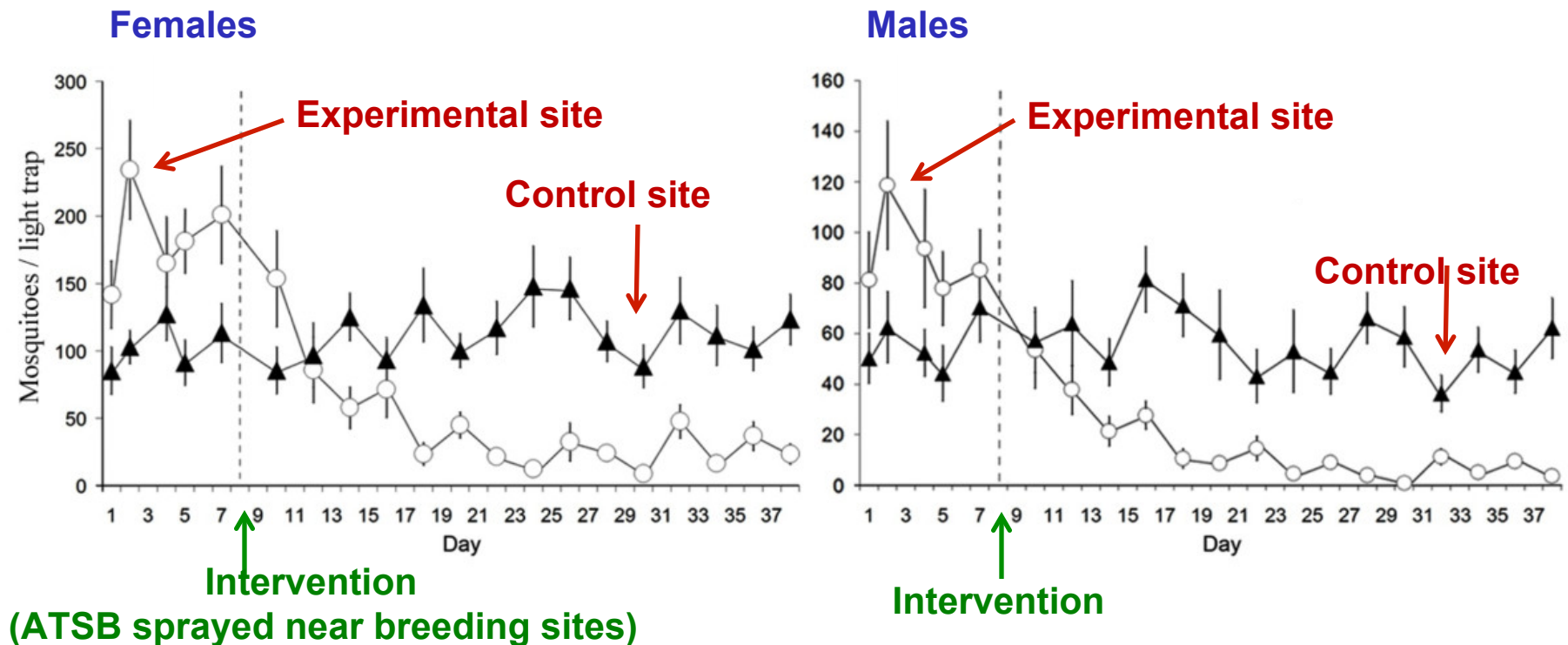
- ***Anopheles sergentii***
- ***Culex pipiens***

- **Recently tested in Mali on:**

- ***Anopheles gambiae***
- **+ more field trials being planned...**

# Successful field trial of attractive toxic sugar bait (ATSB) plant-spraying methods against malaria vectors in the *Anopheles gambiae* complex in Mali, West Africa

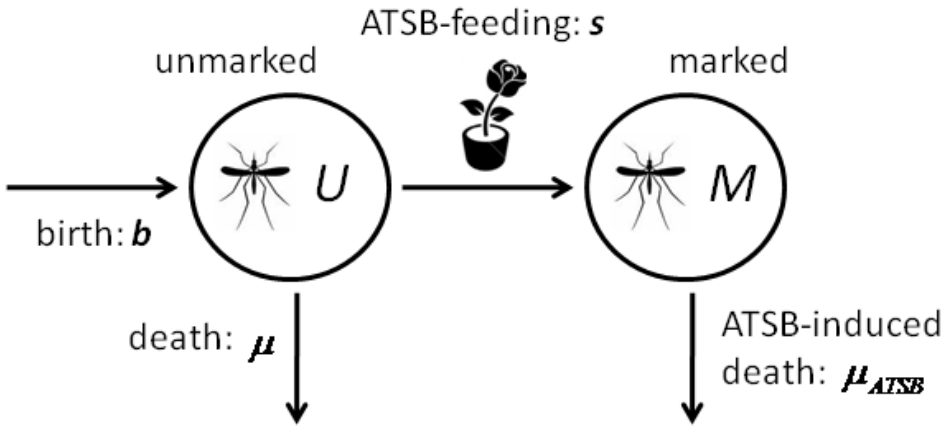
Günter C Müller\*<sup>1</sup>, John C Beier<sup>2</sup>, Sekou F Traore<sup>3</sup>, Mahamoudou B Toure<sup>3</sup>, Mohamed M Traore<sup>3</sup>, Sekou Bah<sup>4</sup>, Seydou Doumbia<sup>3</sup> and Yosef Schlein<sup>1</sup>



- Muller GC, Beier JC, Traore SF, Toure MB, Traore MM, Bah S *et al.* (2010) *Malaria J* 9: 210



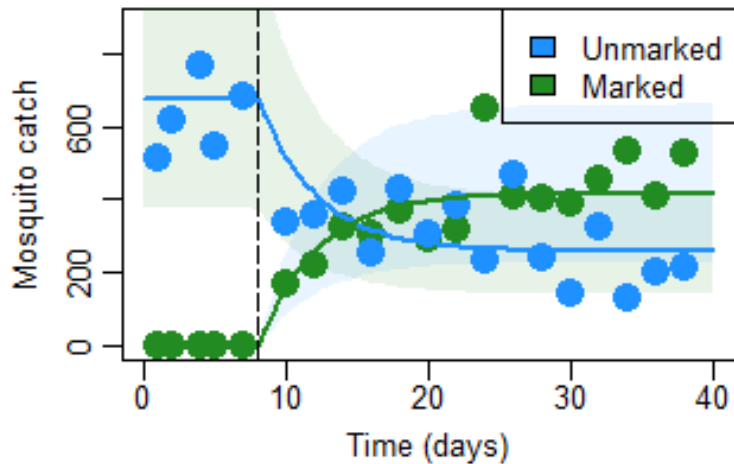
# ATSB data & model fitting



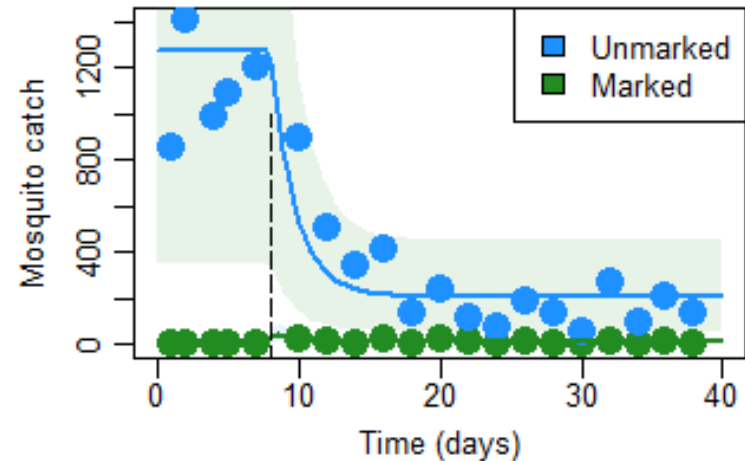
**Rate of feeding on ATSB-sprayed plants:** 0.5 / day  
(CrI: 0.27-0.97)

**Mean lifetime after ingesting of ATSB:** 2.1 hours  
(CrI: 1.1-3.8)

Female control



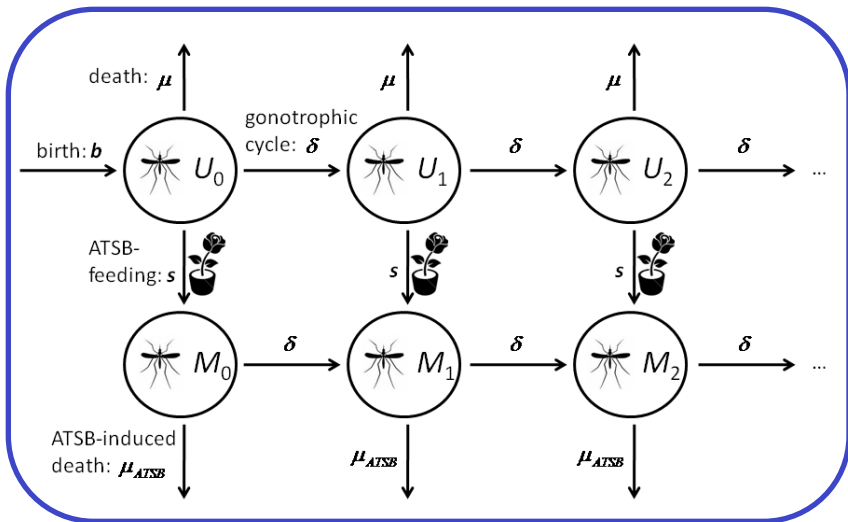
Female experiment



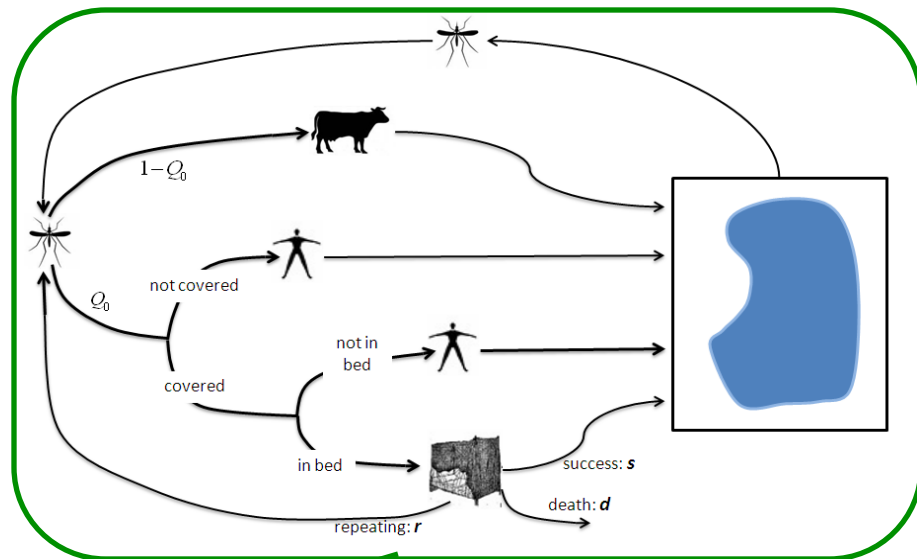
- Marshall JM, White MT, Ghani AC, Schlein Y, Muller GC *et al.* (2013) *Malaria J* 12: 291

# Combine ATSB model with models of mosquito ecology, gonotrophic cycle & vector control

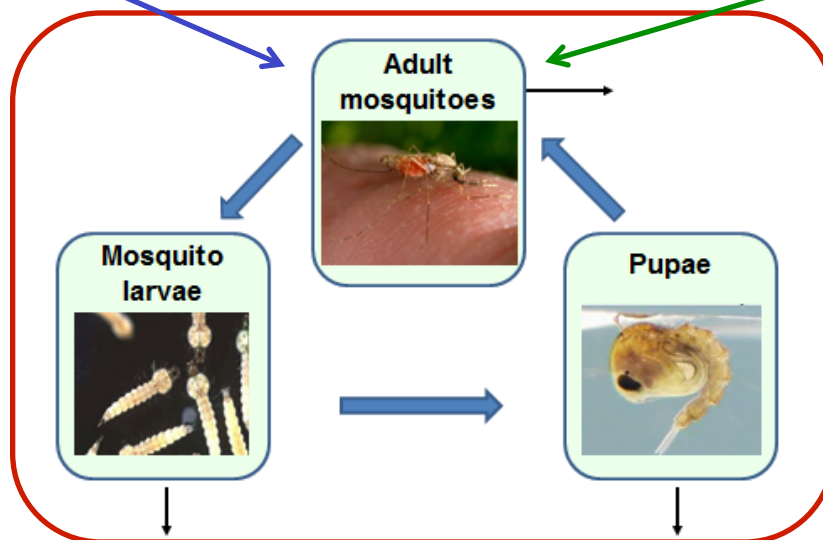
## ATSB model:



## Bed net & insecticide model:

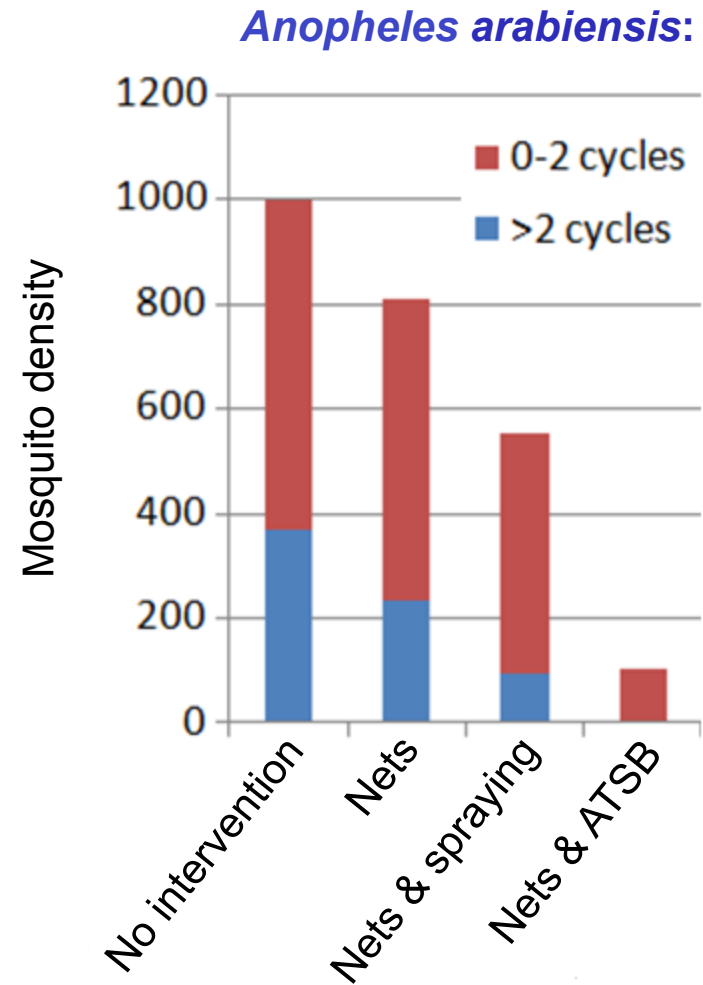
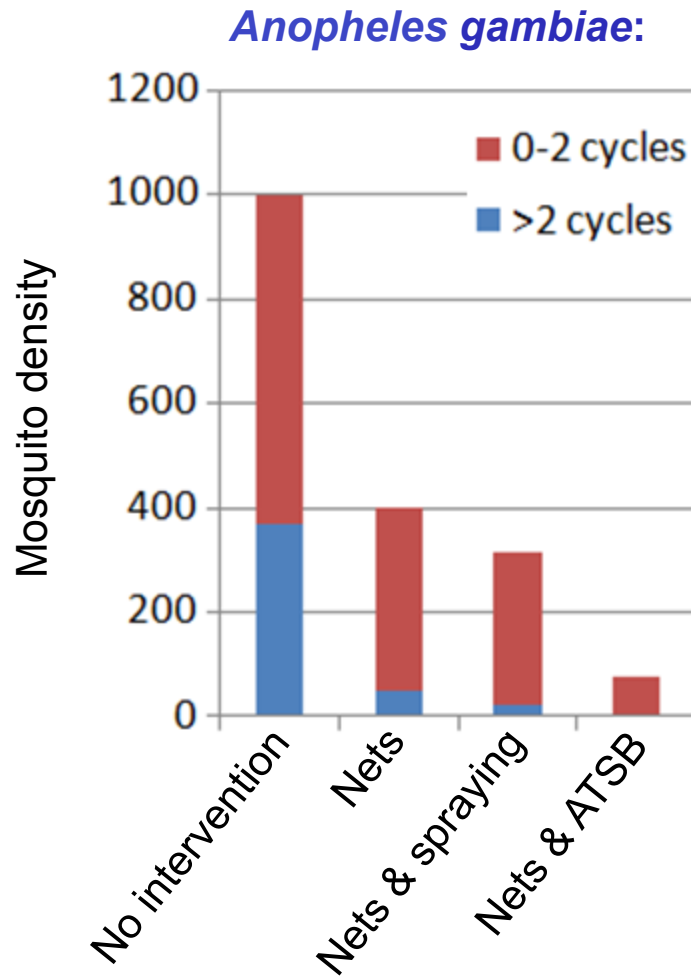


## Mosquito ecology model:



- Marshall JM, White MT, Ghani AC, Schlein Y, Muller GC *et al.* (2013) *Malaria J* 12: 291

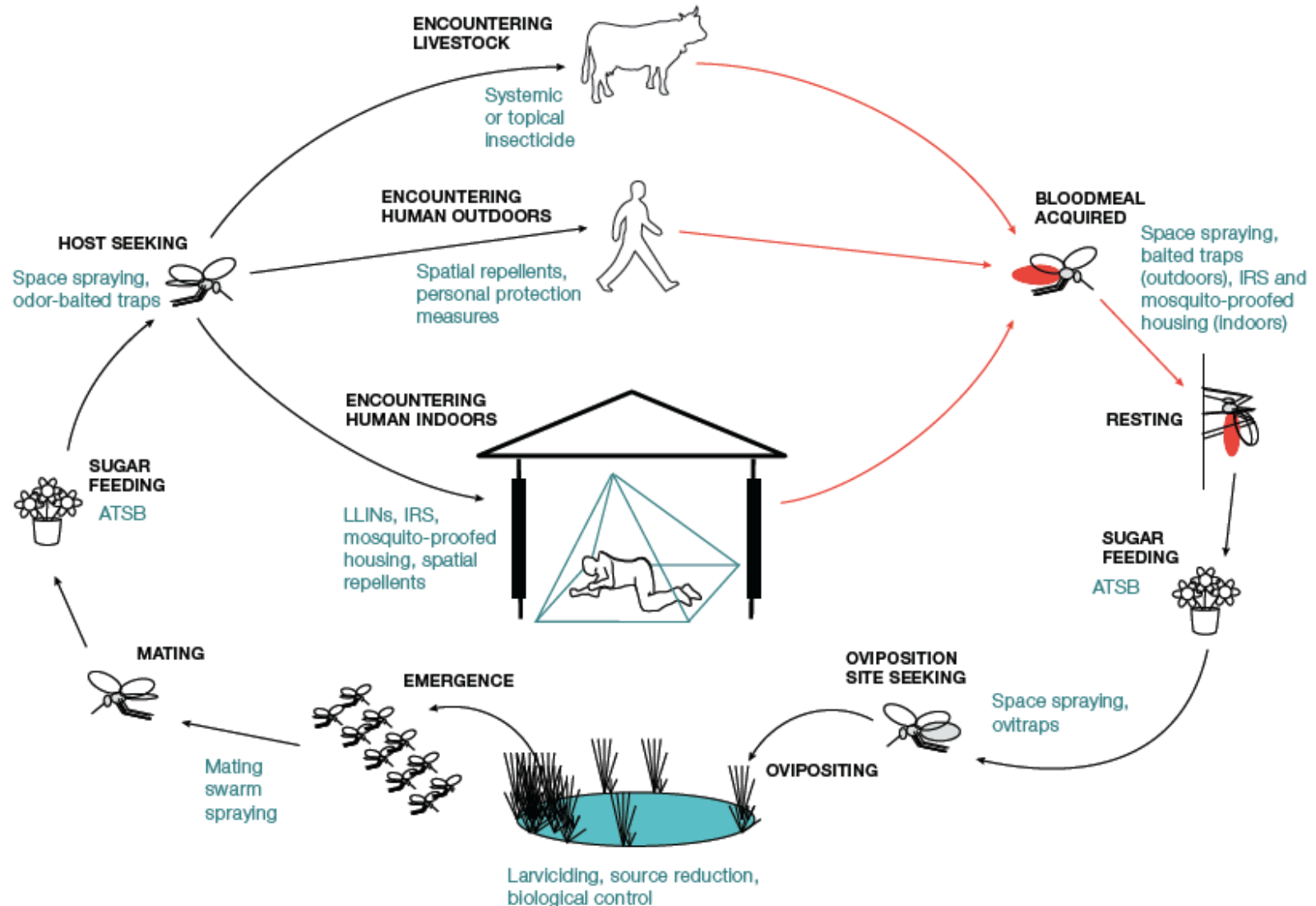
# Predicted combined impact on vector density



- **Nets & spraying significantly reduce *An. gambiae* vector density**
- However, **ATSB complements nets better than insecticide spraying**

- **Nets & spraying are less effective against the exophilic *An. arabiensis***

# Vector Control Optimization Model (VCOM)



- Kiware SS, Chitnis C, Tatarsky A, Wu SL, Sanchez HM *et al.* (2017) PLoS ONE 12: e0187680

# VCOM: Simple

## Vector Control Optimization Model

Home

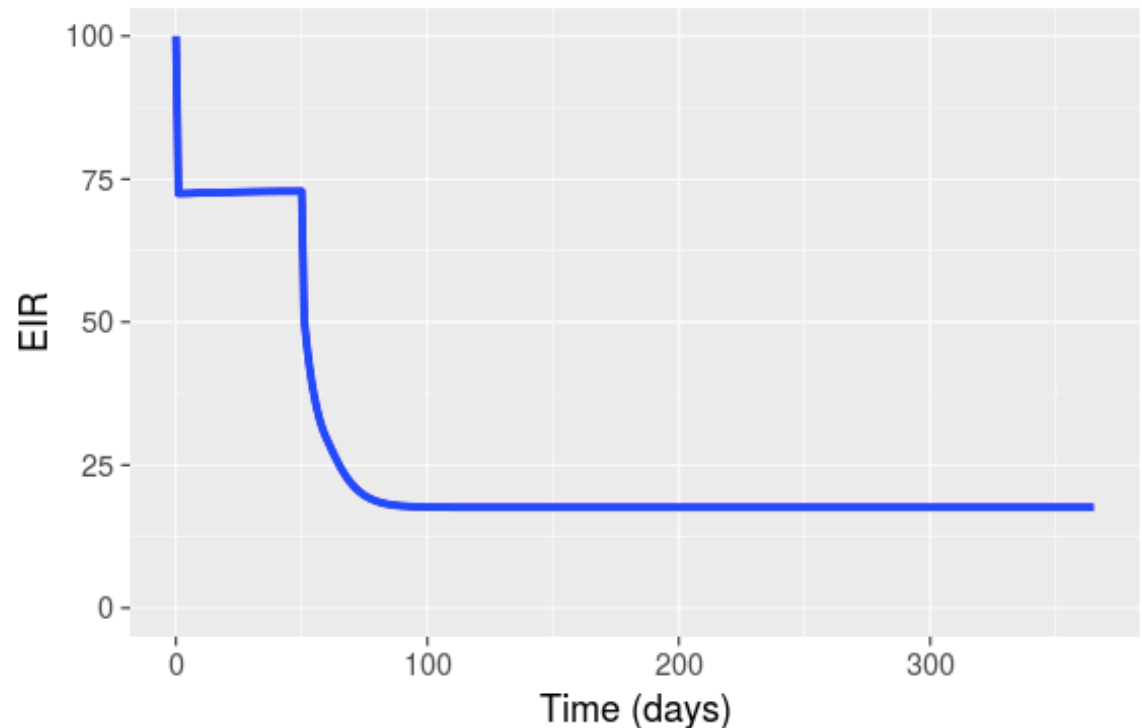
Main

Files Output

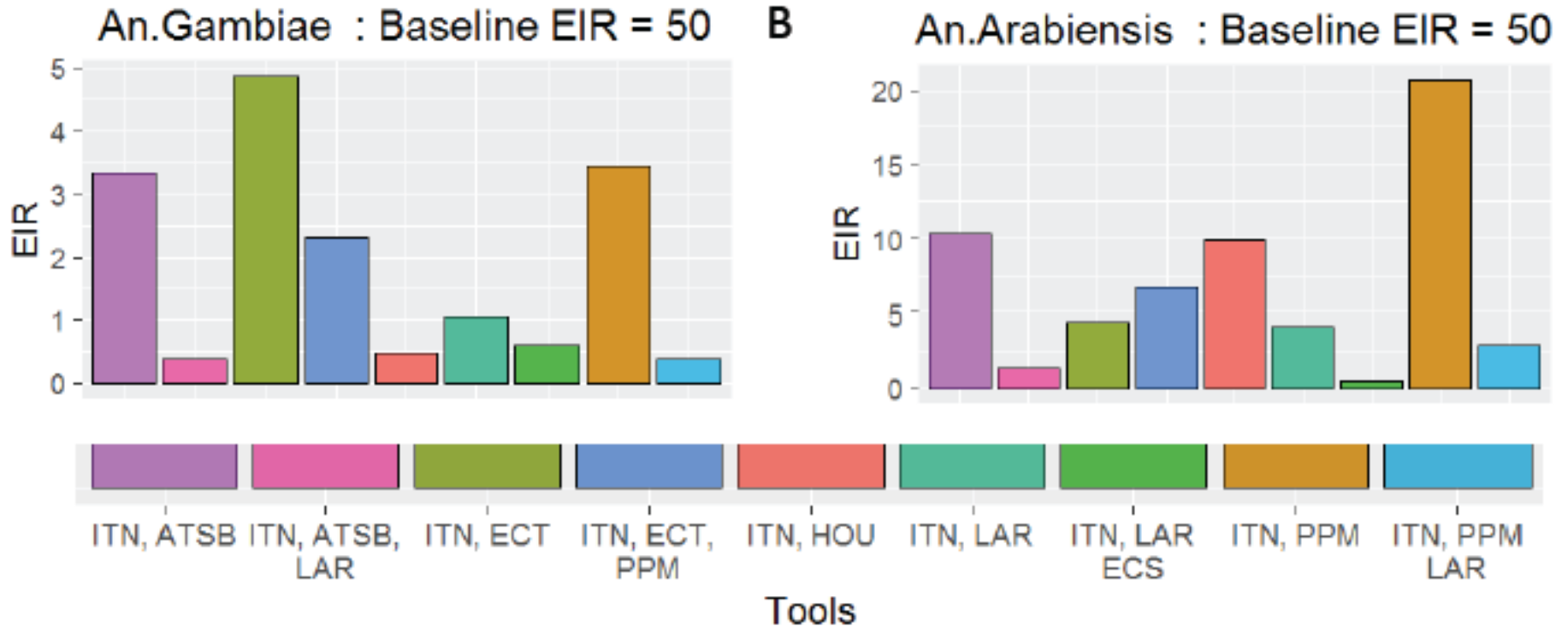
## Instructions

- (1) Select the mosquito species.
- (2) Select the EIR (entomological inoculation rate) level.
- (3) Select the number of days to simulate.
- (4) Run the model! (click the button or hit 'ENTER')
- (5) Setup the desired interventions and repeat step 3 as required.
- (6) Additionally you can download results in the 'Files Output' tab.
- (7) Eliminate malaria!

An. Gambiae Baseline EIR = 100



# VCOM predictions for combined vector control



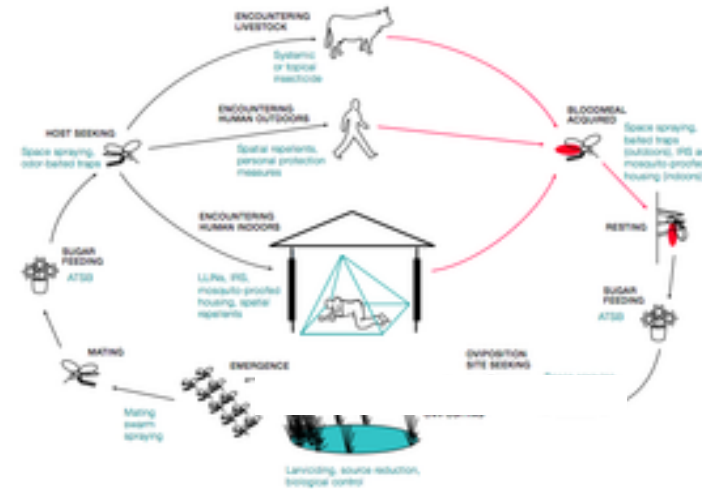
- **Nets, ATSB & larvaciding significantly reduce densities of both *An. gambiae* & *An. arabiensis***
- Other effective combinations for *An. gambiae* are: i) **nets & housing modification**, and ii) **nets, personal protection measures & larvaciding**

- Other effective combinations for *An. arabiensis* are: i) **nets, larvaciding & cattle treated with endectocide systemically**
- **Elimination is very difficult, even under unrealistically optimistic conditions**

# Conclusions

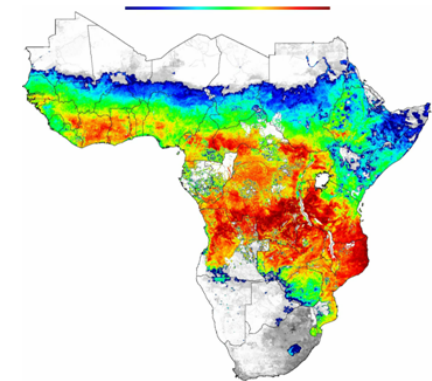
## Q3. Which novel vector control tools should we be prioritizing?

- Attractive toxic sugar baits offer synergies to insecticide-based vector control
- But existing tools are not expected to eliminate malaria in high-prevalence settings



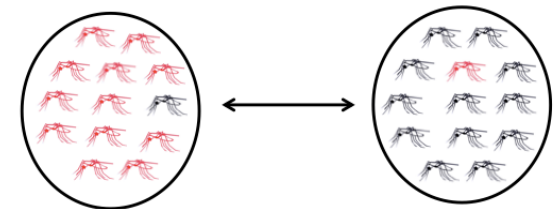
## Q2. Can CRISPR-based gene drive be effective at disease control on a wide scale?

- Multiplexing guide RNAs could sufficiently reduce resistant allele generation rates
- More study is needed of guide RNA multiplexing



## Q1. Is it possible to perform a confined trial of a gene drive system?

- Threshold-dependent systems may be confineable to partially isolated populations
- More study is needed of mosquito population structure



# Acknowledgements

## COLLABORATORS:

### MARSHALL LAB @ UC BERKELEY



Hector M. Sanchez C., Sean L. Wu, Jared Bennett, Samson S. Kiware, Gordana Rasic, Partow Imani, Suzanne Dufault, Biyonka Liang, Tomas Leon.

## OTHERS:

- Akbari & Bier Labs @ UCSD
- James Lab @ UC Irvine
- Hay Lab @ Caltech
- Lanzaro Lab @ UC Davis
- Malaria Elimination Initiative @ UCSF
- Prof David Smith @ IHME, UW
- School of Public Health @ UC Berkeley
- Malaria Modeling Group @ Imperial

## FUNDERS:



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